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Introduction

Alternans is a risk factor for cardiac arrhythmia, including atrial fibrillation.

At the cellular level alternans manifests as beat-to-beat alternations in contraction strength, action potential duration (APD) and magnitude of the Ca transient (CaT).

Electromechanical and CaT alternans are highly correlated and interplay between membrane potential and intracellular Ca cycling plays a key role in the development and stability of cardiac alternans.

In this study we demonstrate that severity of CaT alternans is strongly affected by the shape of APs and investigate if development of alternans can be prevented by targeted modulation of membrane ion channels that determine AP morphology. The findings that CaT alternans can be controlled or even prevented by pharmacologically modulating AP morphology has important ramifications for arrhythmia prevention and anti-arrhythmic therapy strategies.

Methods

Experiments were performed on freshly isolated rabbit atrial cells.

Solutions: External solution (in mM): 135 NaCl, 4 KCl, 2 CaCl₂, 1 MgCl₂, 10 D-glucose, 10 HEPES; pH 7.4 (NaOH). Patch pipette solution (in mM): 130 Kglutamate, 10 KCI, 10 NaCI, 10 HEPES, 0.33 MgCl₂, 4 MgATP; pH 7.3 (KOH).

Electrophysiological measurements. Action potentials were recorded in current-clamp mode (Fig.1). These recorded APs were used as voltage commands in AP voltage-clamp experiments.



Fig.1 A: Simultaneous recordings of AP and [Ca], alternans in current-clamped atrial myocytes. **B:** AP waveforms recorded during large (AP_{CaT Large}) and small (AP_{CaT Small}) alternans CaTs were used as voltage commands to pace cells during AP voltage clamp experiments. C: During AP-clamp experiments trains of voltage stimuli were applied as same-shape $AP_{CaT Large}$, same-shape $AP_{CaT Small}$ and alternans AP stimulation protocols.

[Ca], measurements. Simultaneously with electrophysiological recordings cytosolic [Ca], was recorded using Fluo-4 or Indo-1 pentapotassium salts (100 µM) added to the pipette solution. The degree of CaT alternans was quantified as the alternans ratio (AR), defined as AR=1-CaT_{Small}/CaT_{Large}; where CaT_{Small} and CaT_{Large} are amplitudes of small and large CaTs of an alternating pair of CaTs.

For [Ca]_{sR} measurements myocytes were loaded with the Ca probe Fluo-5N/AM entrapped in sarcoplasmic reticulum.

All single cell experiments were performed at room temperature (22-24°C).

Whole heart. Atrial electrograms were recorded by placing two electrodes on the left atria in Langendorf perfused isolated rabbit hearts. Electrical stimulation was applied to the right atria and stimulation frequency was gradually increased to induce atrial T-wave (T_a) alternans. Experiments were performed at 37°C.

Statistical significance was evaluated using unpaired and paired Student's t-test and differences were considered significant at p < 0.05. Summarized data is presented as mean ± SEM.



alternans compared to AP_{CaT Large} stimuli (n=28). CaT alternans ratio is further increased and the pacing threshold is lowered when alternans AP protocol is applied (n=18).

Action potential shortening prevents atrial calcium alternans



Fig.5 A: K currents recorded in control and in presence of 20 µM NS1643. B: Effect of NS1643 (20 μ M), agonist of hERG K (K_v11.1) channels (**a**), and ML277 (500 nM), agonist of slow delayed rectifier K (K_v7.1) channels (**b**), on action potential morphology in atrial cells paced at 1Hz and exhibiting no CaT alternans.

Fig.7 Effect of K channel agonists NS1643 (A) and ML277 (B) on CaT alternans in currentclamped atrial myocytes. Action potential and [Ca]_i traces observed before and after application of NS1643 or ML277 are shown in panels a. Panels b: CaT alternans ratio in individual cells in control and after the application of K channels agonists. Panels c: mean CaT alternans ratio in control and in presence of K channel agonists



CELLULAR SIGNALING

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