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*Dec 4, 2019*

**Lens as an Osmotic Pump,  
a Bidomain Model**

**DOI: 10.13140/RG.2.2.25046.80966**

Thanks to Weishi and Weizhang for inviting me  
and Gloria for seeing that it all works

**Mathematics Department**

**Kansas University**

**Lawrence KS USA**

## ABSTRACT

The lens of the eye has no blood vessels to interfere with vision. The lens is far too large for diffusion to provide food and clear wastes.

Experimental, theoretical and computational work has shown that the lens supports its own microcirculation. It is an osmotic pump that implements what physiologists have long believed “Convection provides what diffusion cannot.”

We introduce a general (non-electro-neutral) model that describes the steady-state relationships among ion fluxes, water flow and electric field inside cells, and in the narrow extracellular spaces within the lens.

Using asymptotic analysis, we derive a simplified model exploiting the numerical values of physiological parameters. The model reduces to first generation ‘circuit’ models and shows the basis of computer simulations too large to easily understand. The full model helps resolve paradoxes that have perplexed molecular biologists: crucial physiological properties do not depend as expected on the permeability of the lens interior (to water flow).

# **Theme for Recent Lectures at Kansas U**

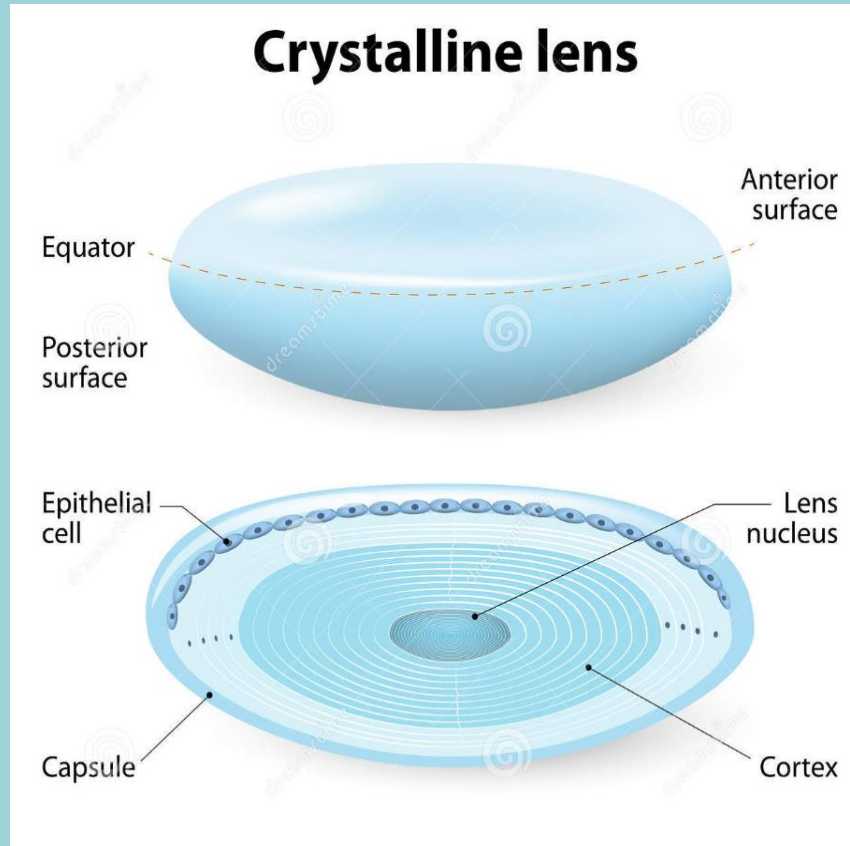
**Bob Eisenberg**

**Examples of Precise Biophysics**  
at different scales, that yield  
**Physical Understanding of Biological Systems**  
and How They Work

**Checked by extensive  
experimentation**

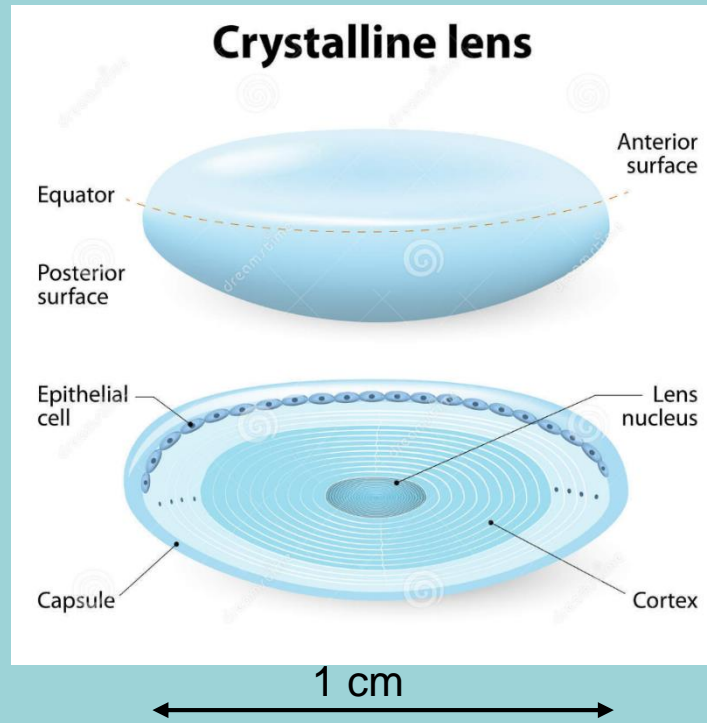
# The Lens of the Eye:

## Bidomain Model of an Osmotic Pump



1 cm

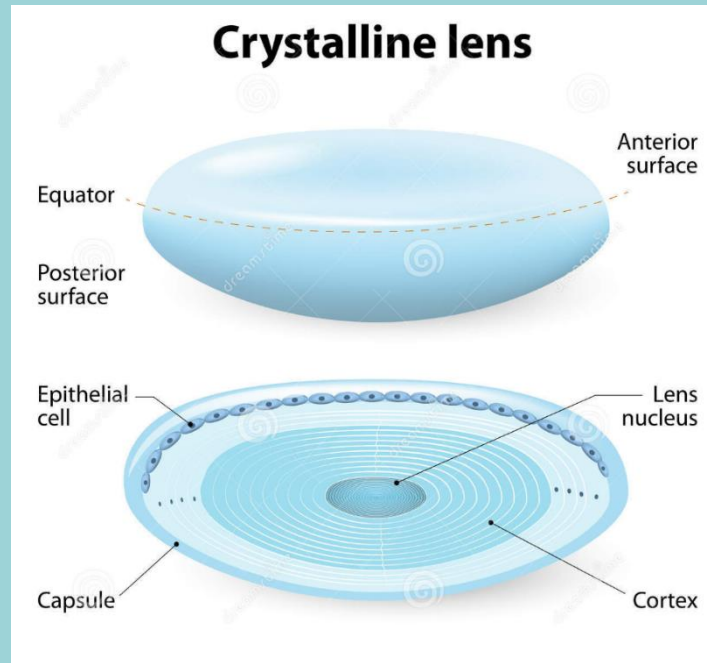




**The Lens is a Device to gather and focus light.**

**It is designed by evolution for a purpose:**

**To gather and refract light**



**The Lens gathers and refracts light.  
No blood vessels!!!**

The Lens is a  
**Preparation**

**Chosen to Study**  
**Convection and Circulation**  
**By an Osmotic Pump**  
Without blood vessels!

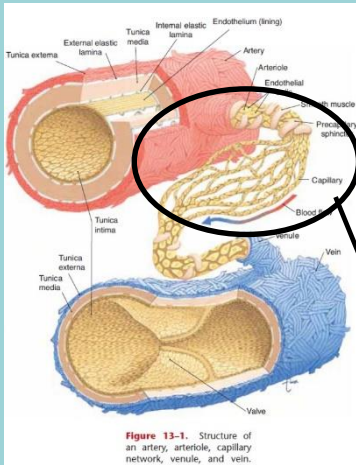
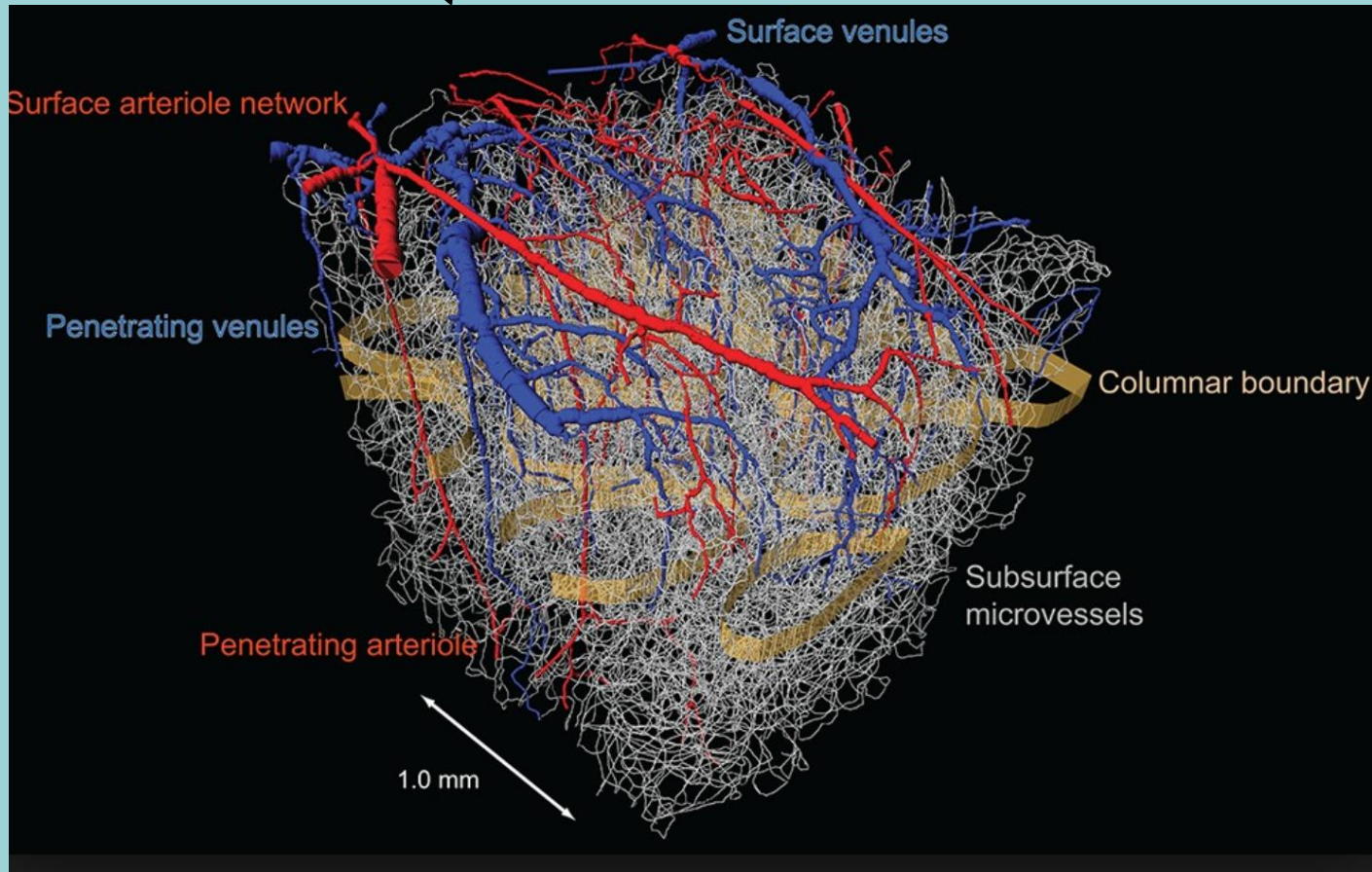


Figure 13-1. Structure of an artery, arteriole, capillary network, venule, and vein.

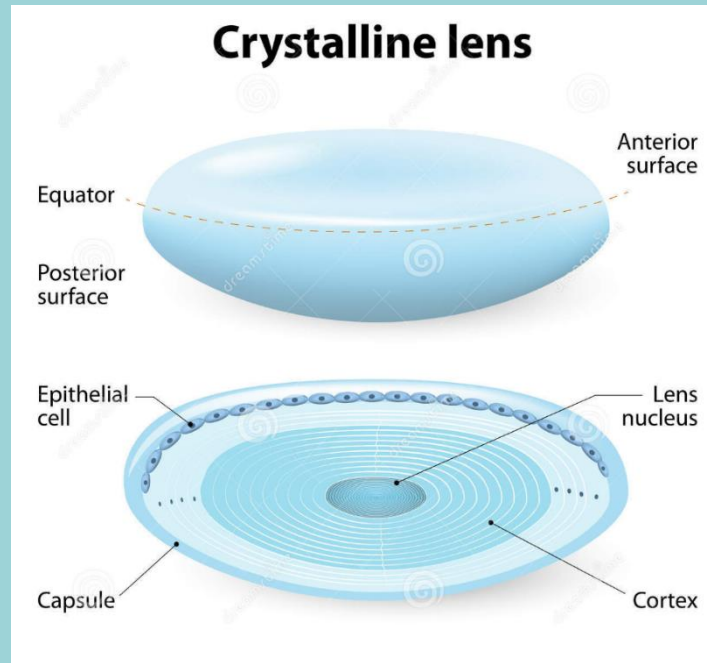
**Circulatory System is Incredibly Dense.  
Convection is needed.**

**Convection delivers what  
Diffusion cannot!**

**Diffusion is too slow to support life!!!**







1 cm

## Central Question:

**How does the lens support circulation  
without blood vessels?**

## Central Idea of Biological Experiments

# Preparation

We imagine that other biological systems  
use the same mechanism as the lens,  
most importantly  
the tiny extracellular spaces in the brain

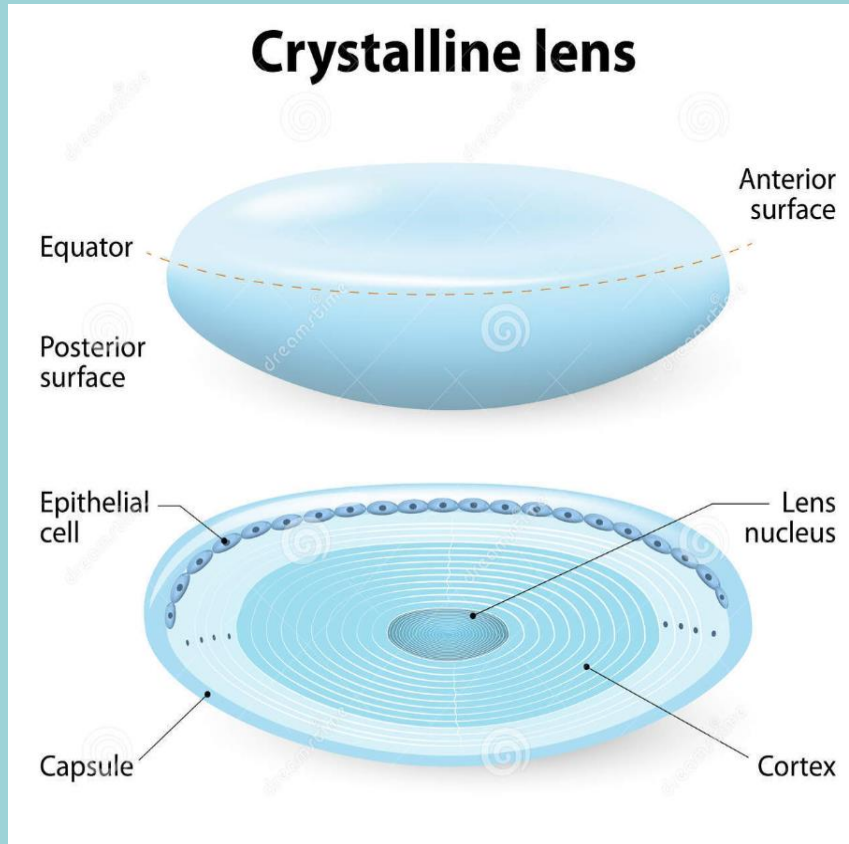
Central Question:

How does the lens support circulation  
without blood vessels?

Central Question:

How does the lens support circulation without blood vessels?

**ANSWER:**  
**The Lens is an**  
**Osmotic Pump**



1 cm



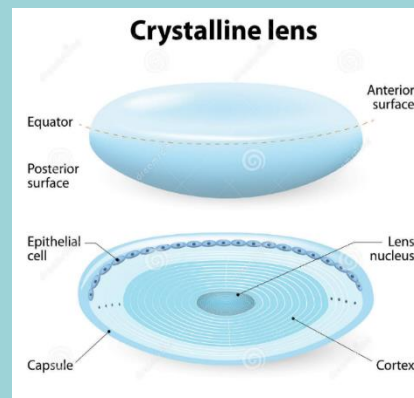
## The Lens

### An Osmotic Pump

Can we prove that the lens is an  
Osmotic Pump?

Are Osmotic Pumps a  
**GENERAL BIOLOGICAL MECHANISM?**

This is a seminar on Quantitative Biology as well as math



1 cm

**Start with the Electrical Structure  
because  
Electrical Methods are Most Precise**

## **Structural Analysis of Electrical Properties**

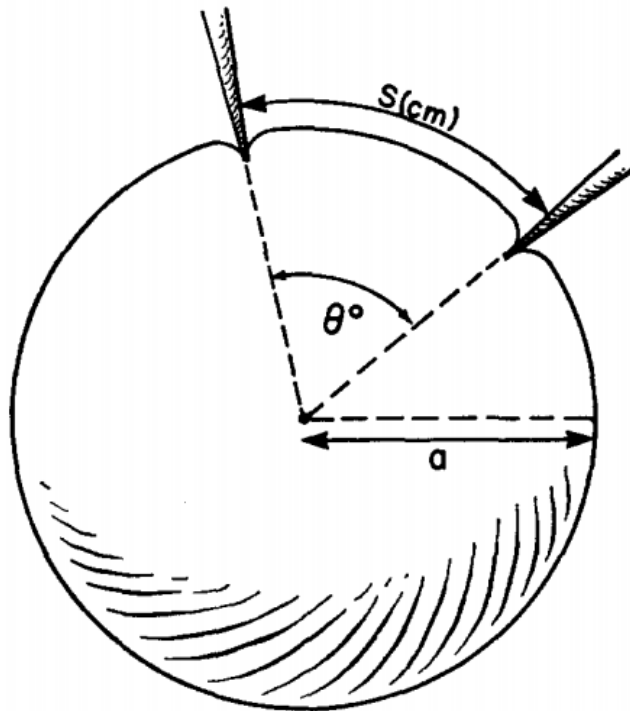
- 1. Qualitative Structure = Anatomy**
- 2. Quantitative Structure Stereology**
- 3. Electrical Model (from Poisson)**
- 4. Impedance Measurements (DC to  $10^4$  Hz)**
- 5. Parameter Estimation**
- 6. Checking Parameter Invariance**
- 7. Prediction of Biological Function**

# Structural Analysis of Electrical Properties uses **MATHEMATICS**

1. Qualitative Structure = Anatomy
2. Quantitative Structure Stereology
3. Electrical Model (from Poisson)
4. Impedance Measurements
5. Parameter Estimation
6. Checking Parameter Invariance
7. Prediction of Biological Function

**INVERSE PROBABILITY  
FIELD THEORY  
CIRCUIT THEORY  
INVERSE PROBLEM  
INVERSE PROBLEM  
FIELD THEORY**

# Electrical Methods are Most Precise Spherical Cell



The steady-state solution can be written as

$$V_m = \frac{i_o R_m}{4\pi a^2} \{ [1 - 2a/\Lambda] \{ 1 + (a/\Lambda)D - (a/\Lambda)^2 E_o \} + (a/\Lambda) \csc \theta/2 \} \quad (1)$$

where

$$D = \ln \frac{\csc^2 \theta/2}{1 + \csc \theta/2} \quad (2)$$

$$E_o = \sum_{n=1}^{\infty} \frac{P_n(\cos \theta)}{n^2} \quad n = 1, 2, 3, \dots$$

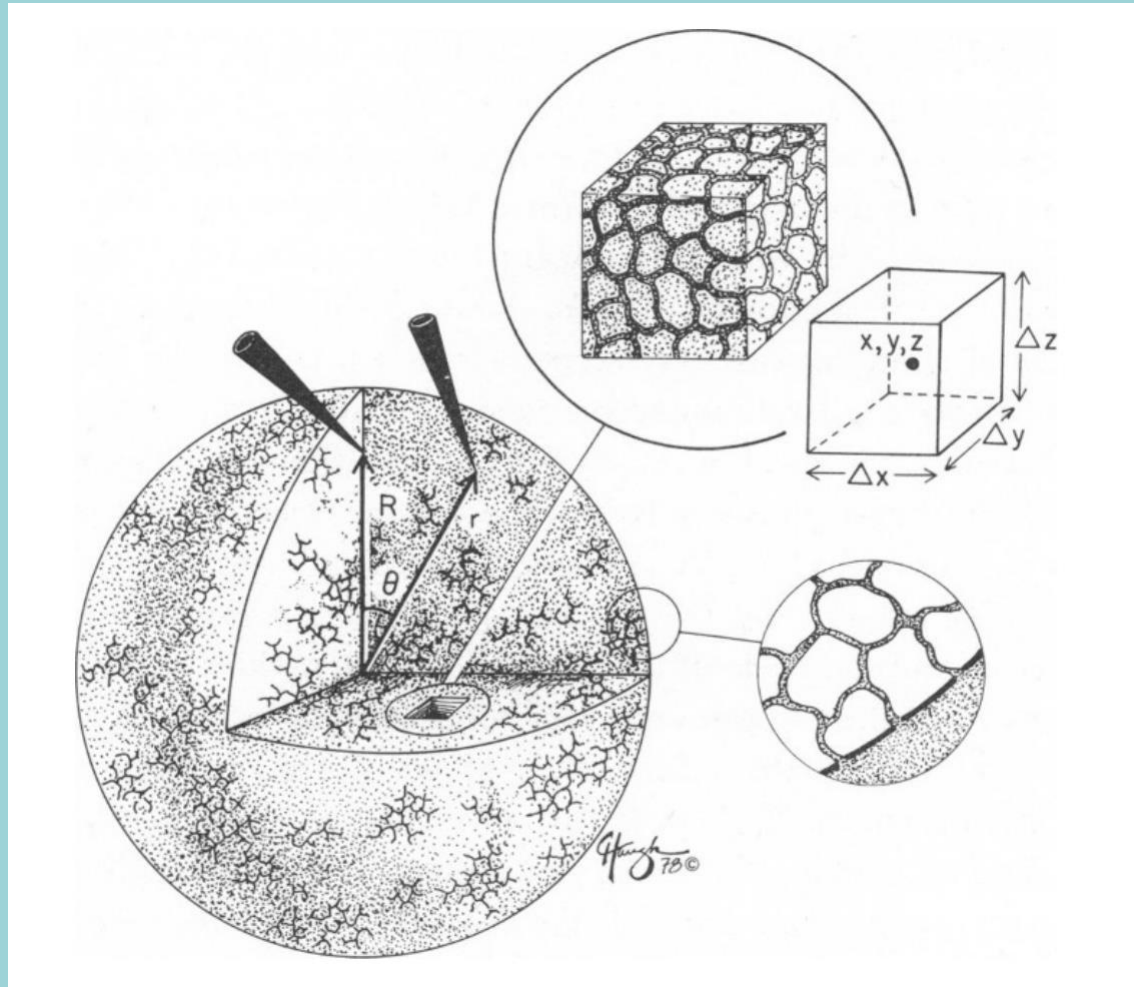
$$\frac{V_m}{i_o} \approx \frac{R_m}{4\pi a^2} \{ 1 + (a/\Lambda) \csc \theta/2 \} = \frac{R_m}{4\pi a^2} + \frac{R_i}{4\pi a} \csc \theta/2 \quad (3)$$

## Electrical Models often using Singular Perturbation Theory

exploiting membrane boundary condition

1. *J Gen Physiol*, 1970. **55(6)**: p. 736-57.
2. *Prog. Biophys. Mol. Biol.*, 1970. **20**: p. 1-65
3. *SIAM J. Appl. Math.*, 1971. **21(2)**: p. 339-354.
4. *Biophys J*, 1972. **12(4)**: p. 384-403.
5. *Annu Rev Biophys Bioeng*, 1973. **2**: p. 65-79.
6. *J. Mathematical Biology*, 1975. **2**: p. 277-300.
7. *J. Mathematical Biology*, 1975. **2(4)**: p. 301-316.
8. *SIAM J. Appl. Math.*, 1976. **30(2)**: p. 222-239.
9. *Ann N Y Acad Sci*, 1977. **303**: p. 342-54.
10. *Biophys J*, 1977. **17(1)**: p. 57-93.
11. *Biophys J*, 1978. **23(2)**: p. 277-84.
12. *CRC Crit Rev Bioeng*, 1980. **4(3)**: p. 203-32.
13. *Biophys J*, 1983. **44(2)**: p. 225-48.
14. *Biophys J*, 1983. **42(1)**: p. 55-9.
15. "Membranes, Channels, and Noise 1984", p. 49-116.
16. *Biophys J*, 1985. **48(2)**: p. 253-67.
17. **Kevorkian, J. and J.D. Cole, 1996**  
***Multiple Scale & Singular Perturbation Methods.***

# Electrical Models of Syncytia Can Be Derived Using Definitions of Vector Operators, Div and Grad



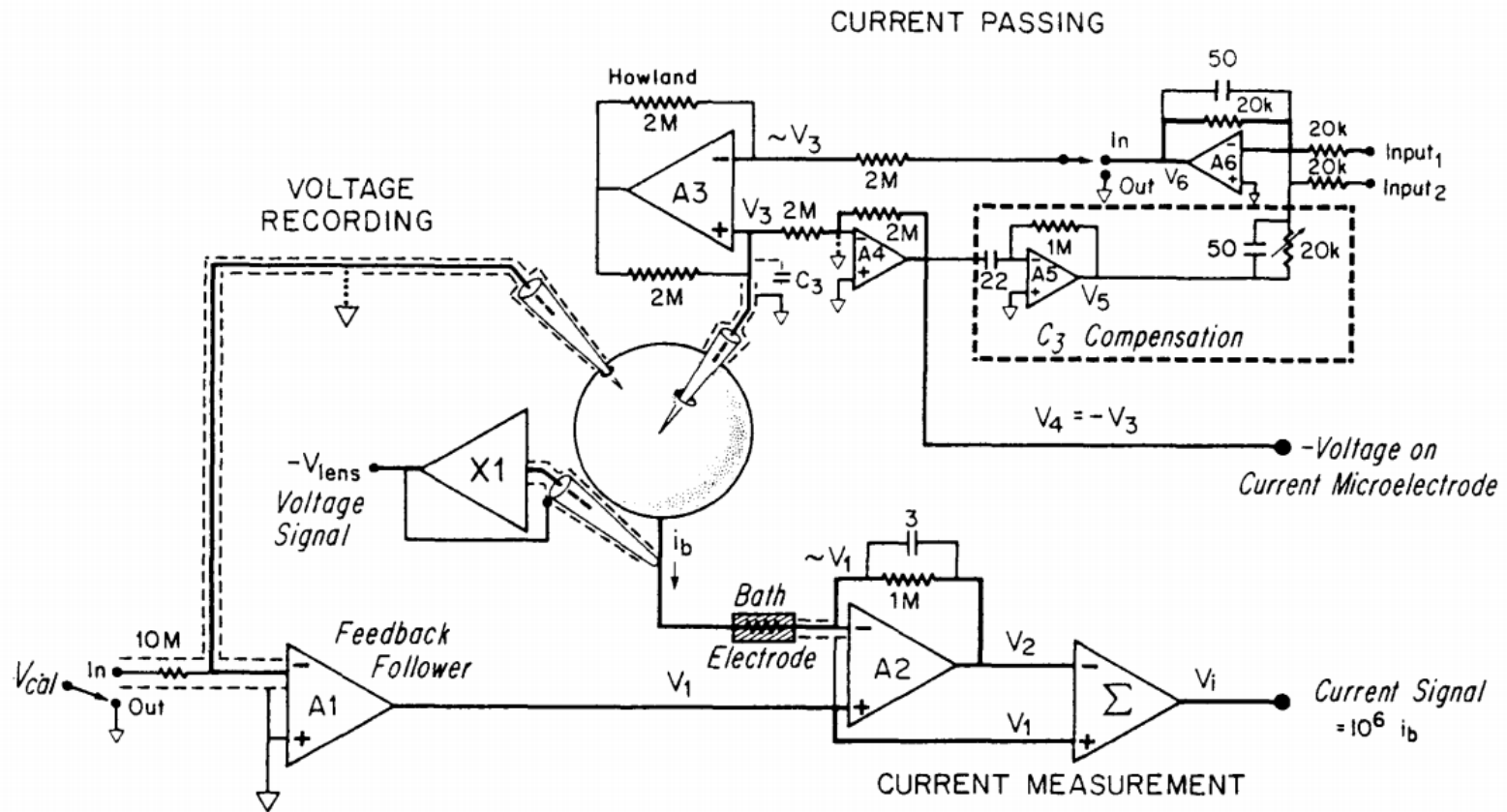
Eisenberg, Barcion, and Mathias. *Biophys. J.* 25: 151-180 (1979).



The Lens: An Osmotic Pump

# Electrical Methods are Most Precise

## Impedance Measurements of Spherical Syncytia



Mathias, Rae, and Eisenberg. Biophys. J. 34: 61-85 (1981).

# Electrical Measurements are Precise

## Spherical Syncytia

### TYPICAL DATA not selected for fit

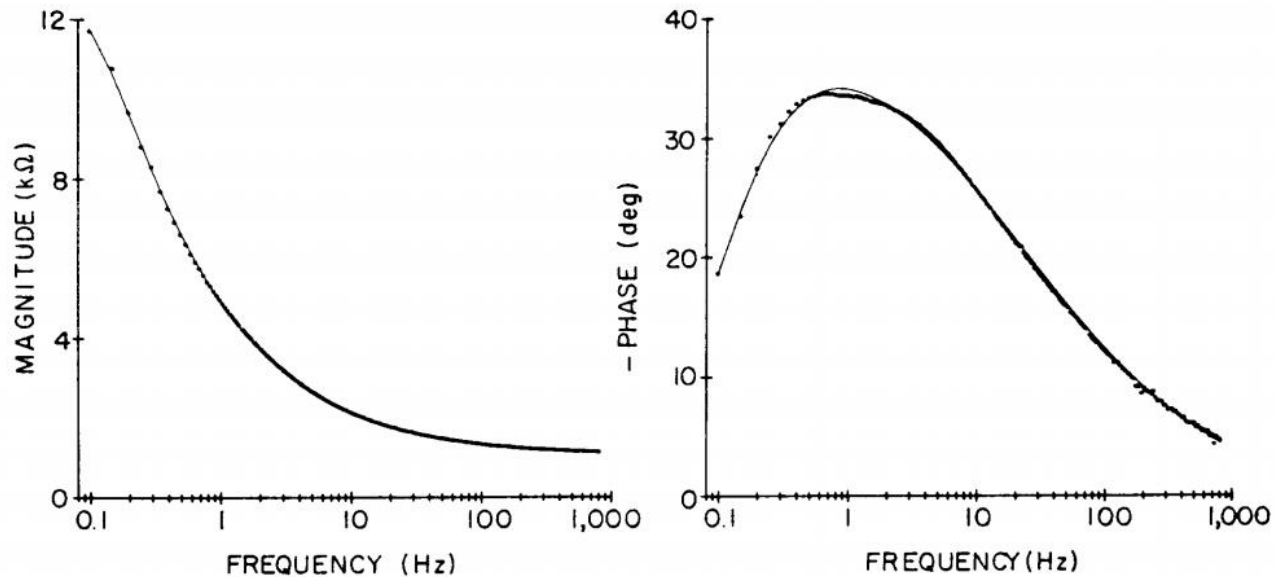


FIGURE 9 The magnitude, phase angle, and theoretical fit to impedance data of lens 2-8 taken from one electrode location. The electrode was at  $r_1 = 0.93$  mm in a lens of radius  $a = 1.33$  mm. The circuit parameters determined from the outermost location of the electrode are quite close to the average parameters determined from many locations (Table IV).

Rae, Mathias, and Eisenberg. *Exp. Eye Res.* 35: 471-490 (1982).

# How is this precision possible?

## Biology has Invariants!

Cell Interior is a (nearly) pure resistor  
Membrane is a (nearly) a pure capacitor

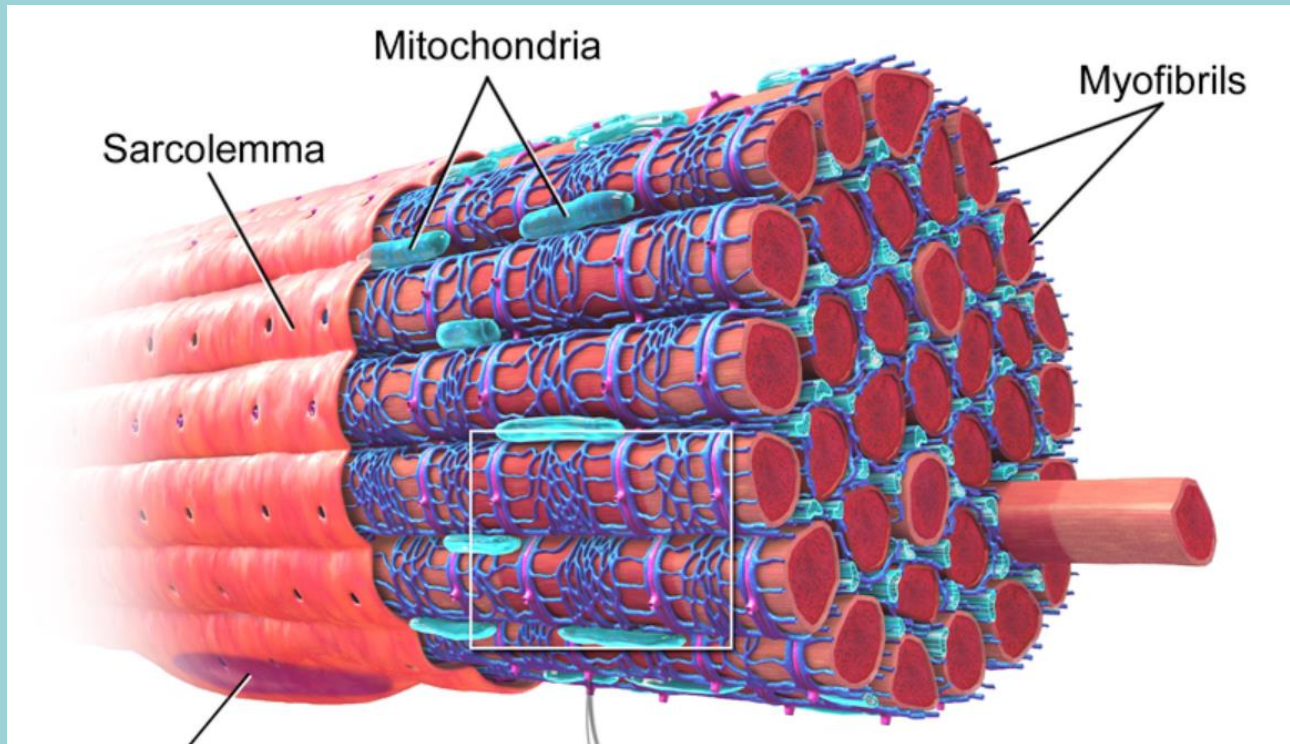
Biology has Invariants!

**Cell Interior  
is a (nearly) pure resistor**

$$i = \frac{V}{R}$$

How is this precision possible?  
Biology has Invariants!

**Interior of Cells is (nearly) a pure resistor**  
*even a muscle fiber*

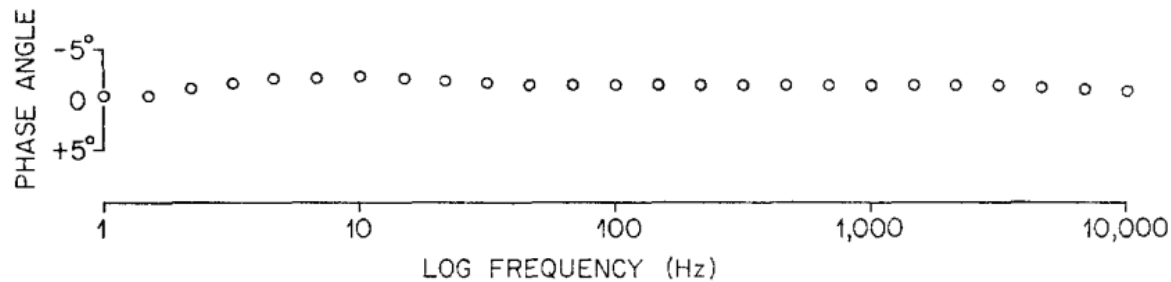
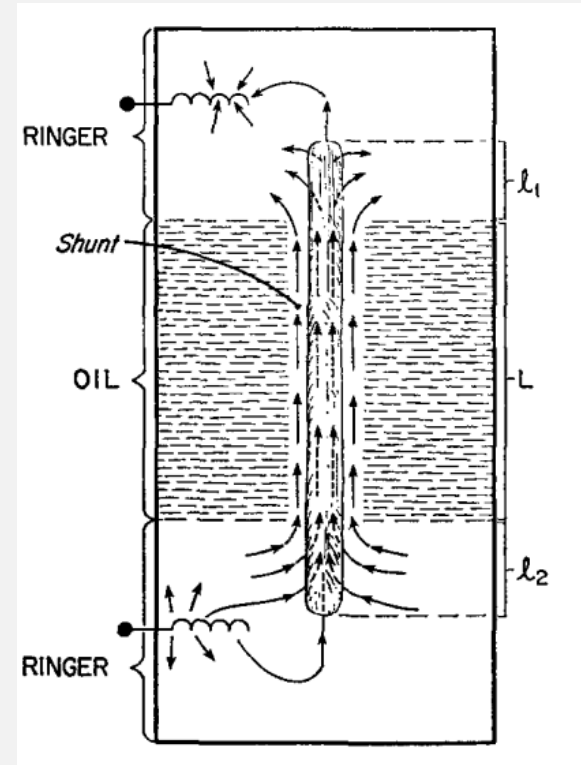


# Muscle Interior is a Pure Resistor

## Biology has Invariants

TABLE I  
RESULTS FROM 14 MUSCLE FIBERS

	$\phi_{\max}$
	<i>degrees</i>
	-1.48
	1.33
	0.74
	-2.45
	-2.42
	-2.22
	-1.58
	-1.60
	0.59
	0.68
	-1.79
	-0.62
	-1.86
	-2.25
Mean	-1.07
SD	1.34



**Interior of Cells is (nearly) a pure resistor**  
*even a muscle fiber*

**How is this possible?**

**Electric Current Flows AROUND Obstacles**

**Electric Current Flows through Low  
Resistance Path**

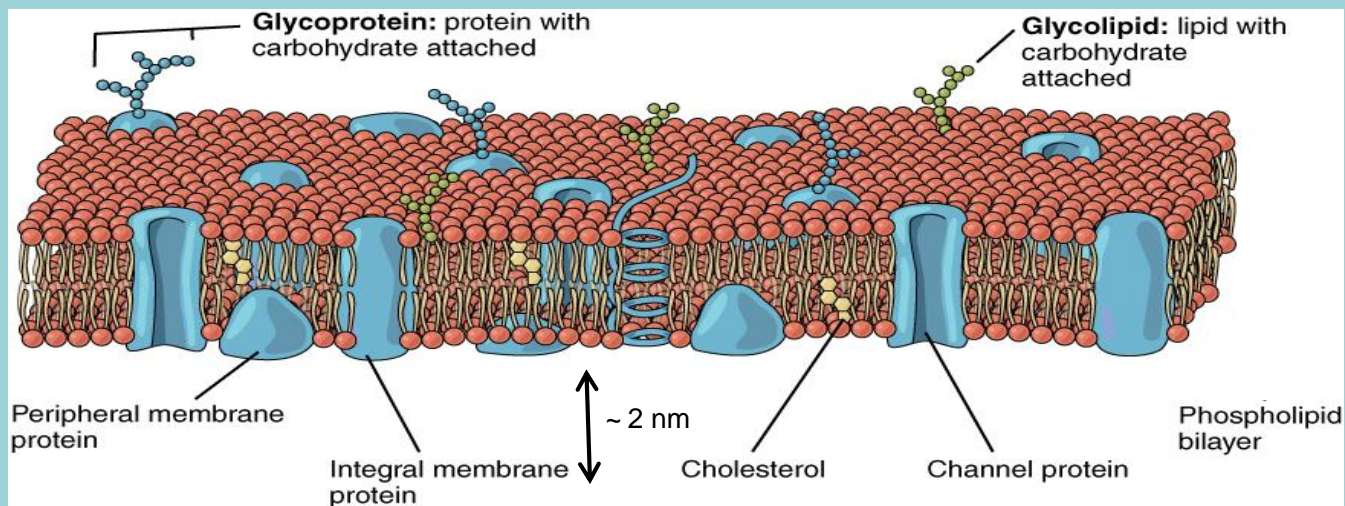
**Property of PARALLEL RESISTORS**

Biology has Invariants!

**Cell Membrane  
is a (nearly) pure capacitor**

$$i = C \frac{\partial V}{\partial t}$$



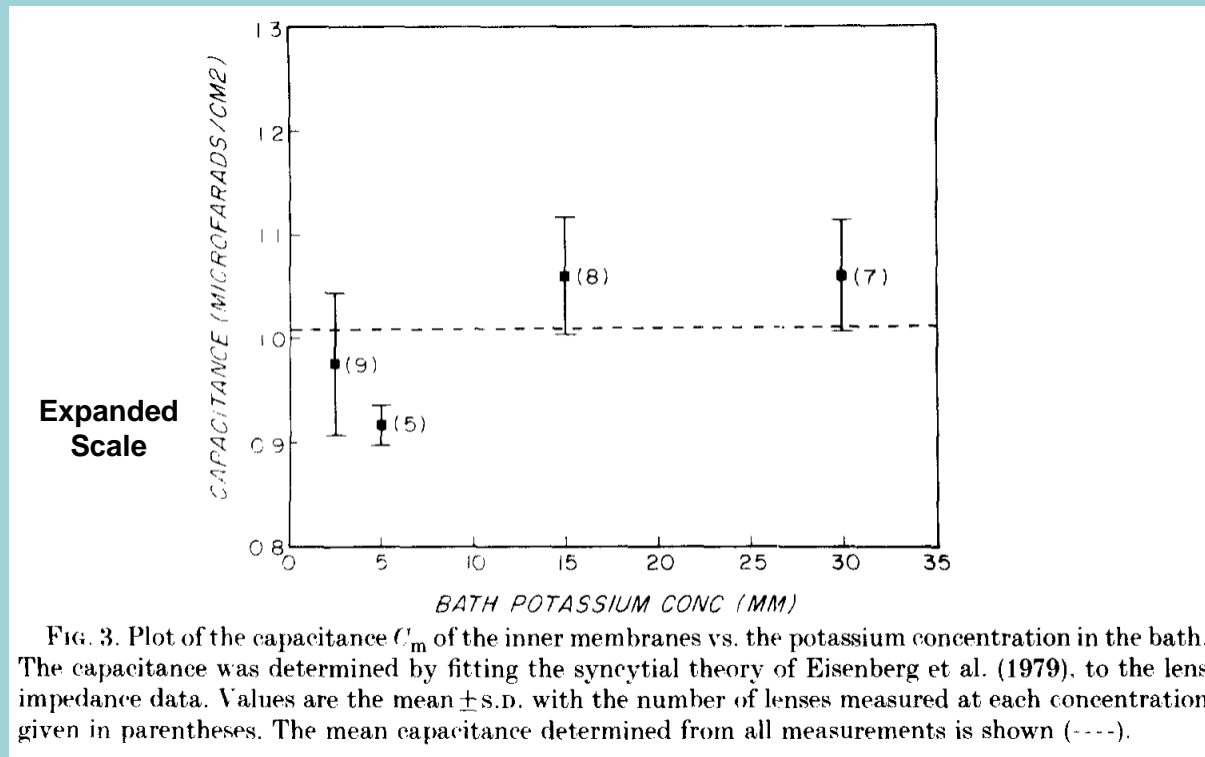


## Denis Haydon Life Work

### Membrane is a (nearly) a pure capacitor

1. *J Theor Biol*, 1965. 9(3): p. 433-43.
2. *J Theor Biol*, 1965. 9(3): p. 422-32.
3. *J Theor Biol*, 1965. 9(2): p. 278-96.
4. *J Gen Physiol*, 1965. 48: p. SUPPL:59-63.
5. *J Mol Biol*, 1968. 32(1): p. 149-50.
6. *Br Med Bull*, 1968. 24(2): p. 124-6.
7. *J Theor Biol*, 1968. 18(3): p. 371-9.
8. *J Am Oil Chem Soc*, 1968. 45(4): p. 230-40.
9. *J Membr Biol*, 1971. 5(3): p. 277-96.
10. *Biochim Biophys Acta*, 1979. 557(1): p. 259-63.
11. *J Physiol*, 1979. 287: p. 38P-39P.
12. *J Physiol*, 1979. 287: p. 2P.
13. *Biophys J*, 1980. 30(1): p. 129-36.
14. *J Physiol*, 1980. 309: p. 229-45.

# Estimated\* Membrane Capacitance of Lens does NOT vary with ion concentrations



\*Note: this estimation involves structure, qualitative and quantitative, impedance measurement, field theory, and solving inverse problem. Errors in any of these will produce estimates that vary with concentration.

The Lens  
An Osmotic Pump

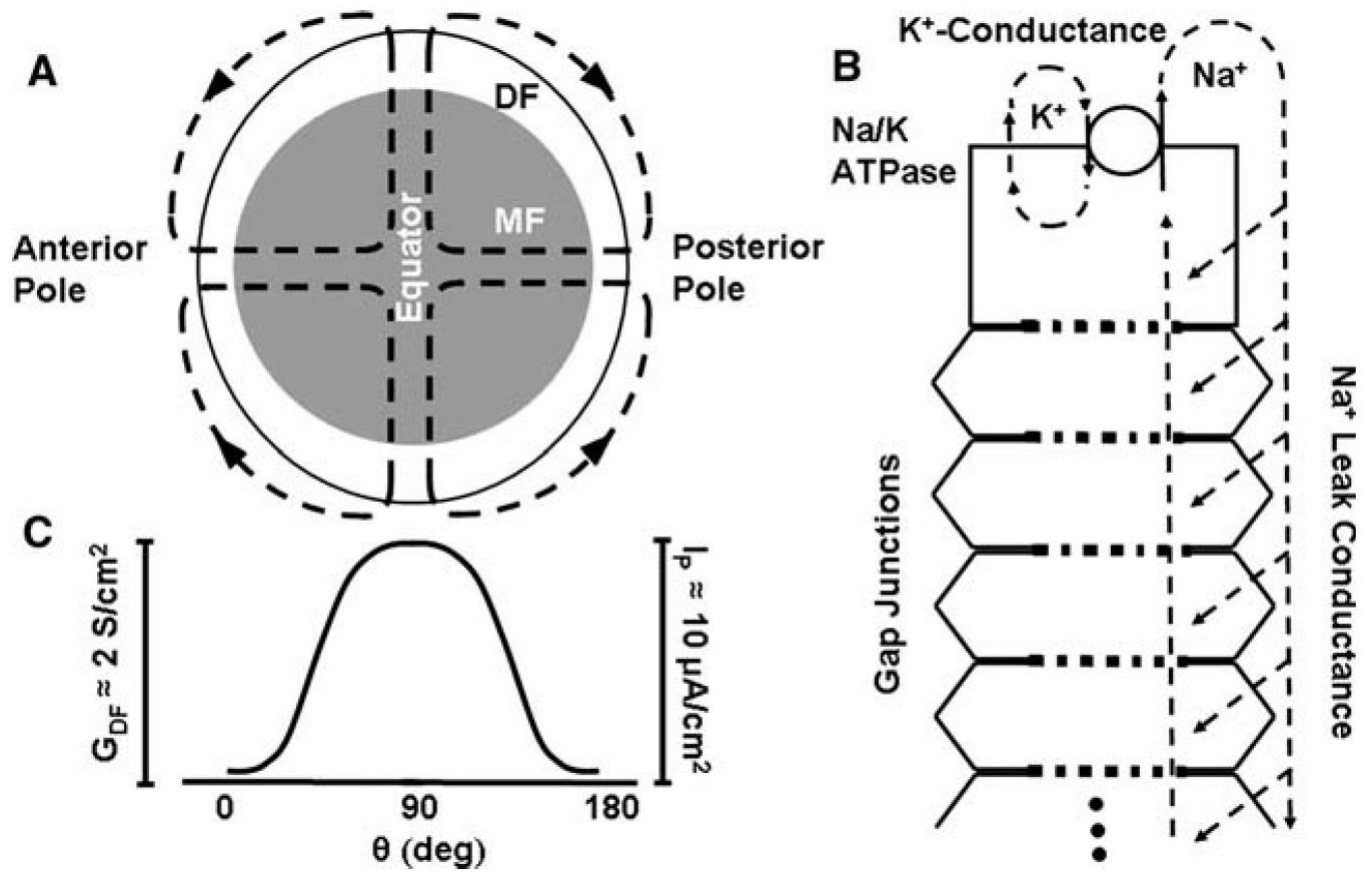
Can we prove that the lens is an Osmotic Pump?

Life work of Richard (Rick) Mathias



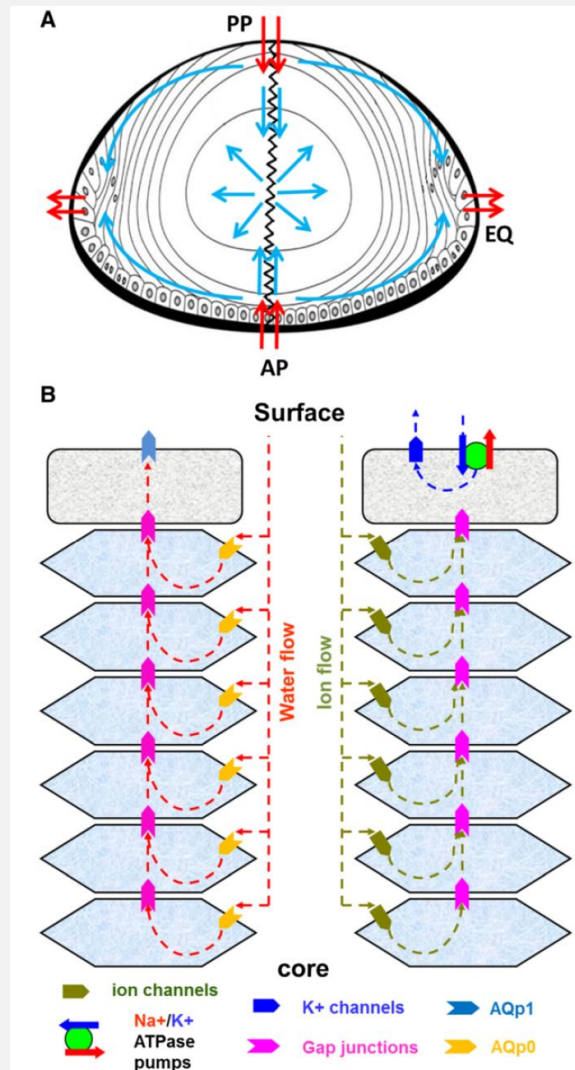
State University of New York SUNY Stony Brook

Too many papers to list! Do Google Search



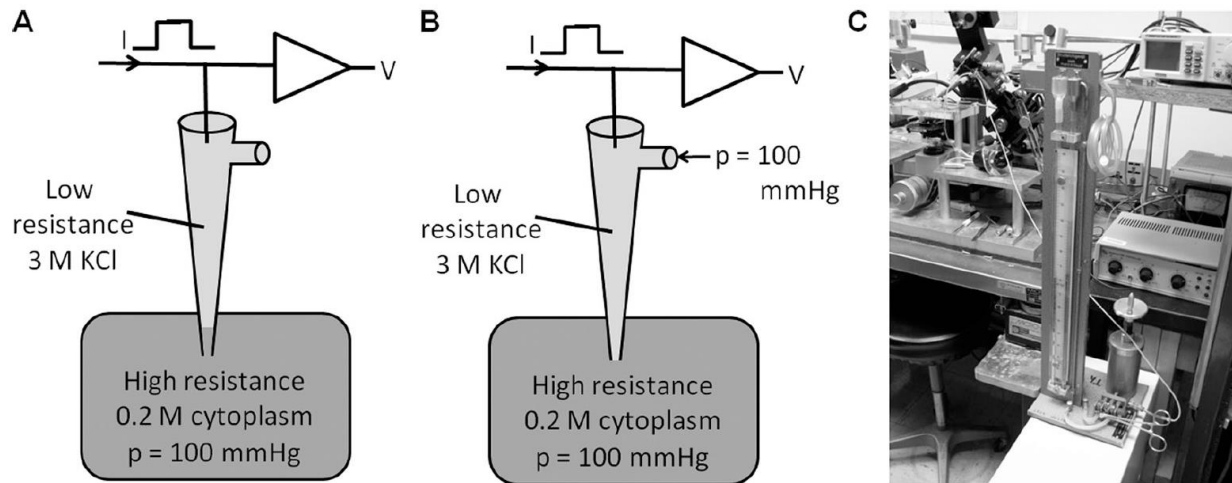
Mathias, Kistler, and Donaldson,  
 J Membr Biol, 2007. 216(1): p. 1-16.

# Computer Model of Lens



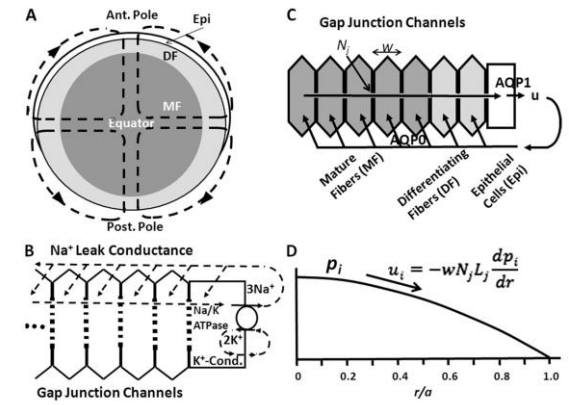
Vaghefi, E., N. Liu, and P.J. Donaldson, *Biomed Eng Online*, 2013. **12: p.85**

# Direct Measurement of Pressure in the Lens



**Figure 2.** The intracellular pressure measuring system. (A) A sketch of the effect of intracellular pressure on the electrode–intracellular solution interface. An elevated intracellular pressure (e.g., 100 mmHg) over that in the bathing solution causes the interface to move up the shank of the electrode, thus filling the narrow tip with relatively high resistance cytoplasm. (B) When the same 100-mmHg pressure is applied to the port of the microelectrode, the interface moves back to the tip as shown, and the electrode resistance is restored to its relatively lower value recorded in the bathing solution. (C) The manometer used to adjust the hydrostatic pressure at the electrode port. The crank drives a piston to create the pressure, which is connected to the electrode port through the plastic tubing. When the electrode resistance is restored to its value in the bathing solution, such that an increase in applied pressure has no effect on resistance, whereas a small reduction in pressure causes a small increase in resistance, we assume the applied pressure equals the intracellular pressure. The value of applied pressure is then read from the column of mercury.

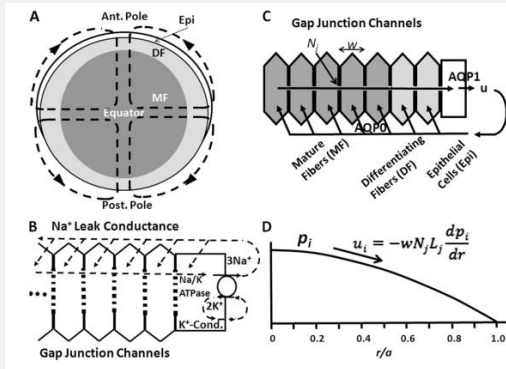
**Gao, Sun, Moore, White, Brink, and Mathias,  
*J Gen Physiol*, 2011. 137: 507-20.**





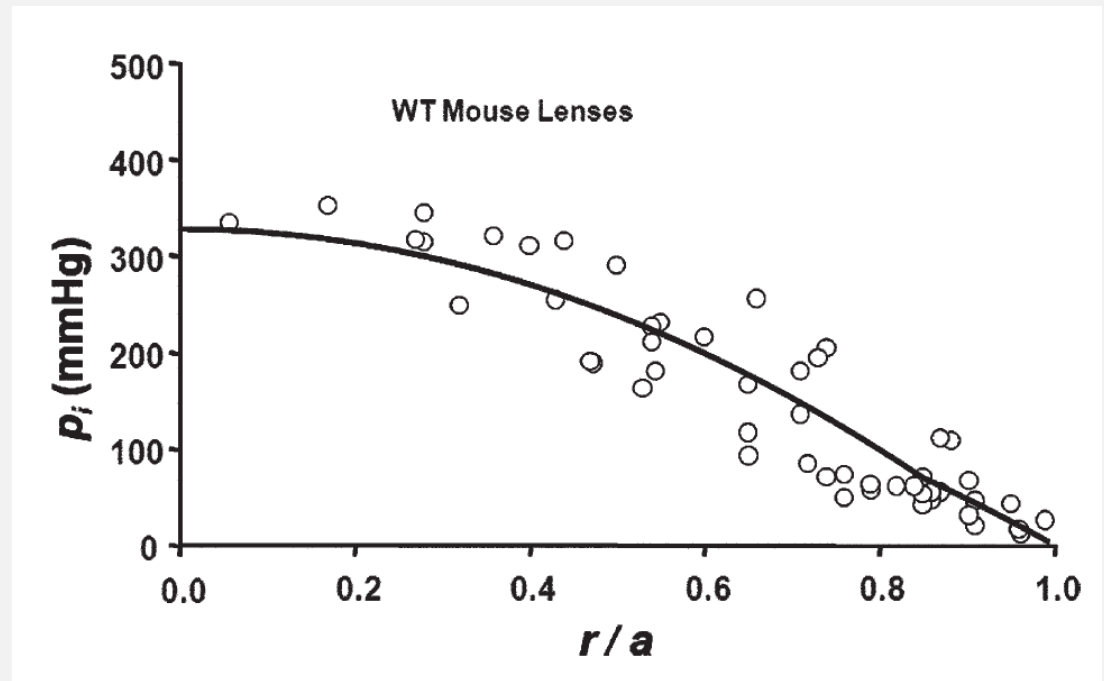
# Direct Measurement of Pressure in the Lens

## Theory

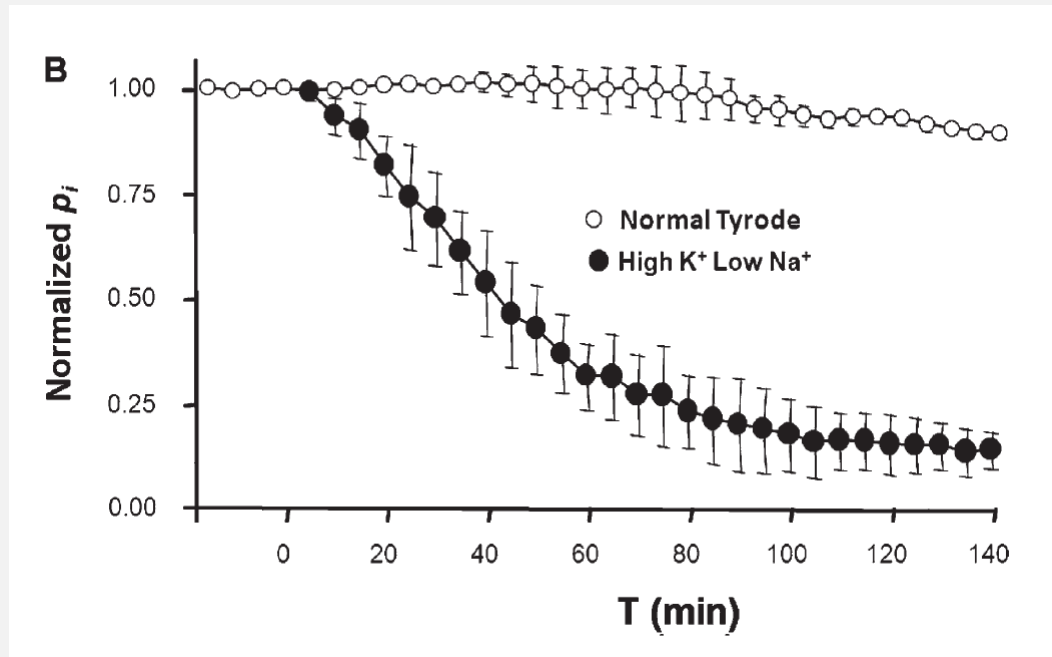


## Experiment

Gao, Sun, Moore, White, Brink, Mathias  
*J Gen Physiol*, 2011  
 137: 507-20.



# Pressure with Reduced Transport in the Lens



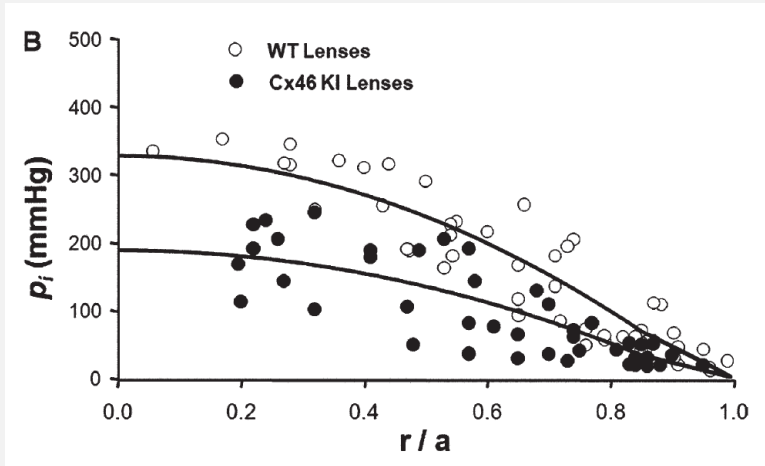
Normal Transport

Reduced Transport

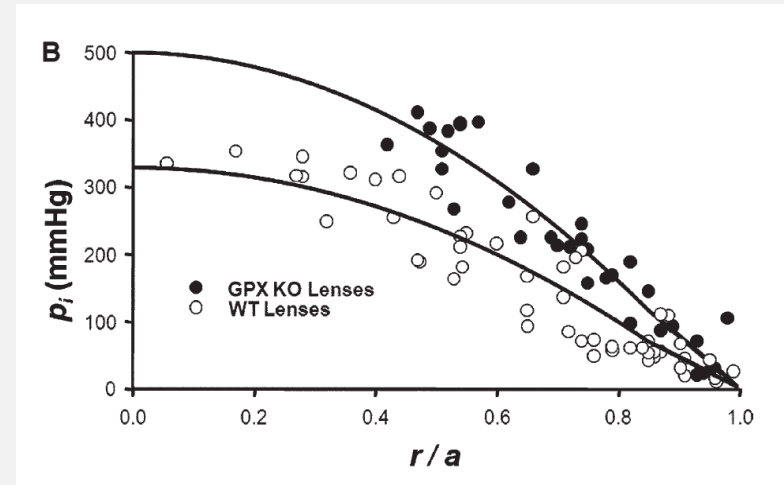
Gao, Sun, Moore, White, Brink, Mathias  
*J Gen Physiol*, 2011  
137: 507-20.



# Changing Cell to Cell Coupling by Molecular Genetics



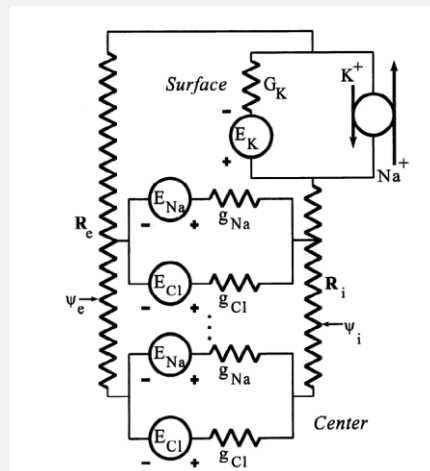
Increasing the Number  
of Junction Proteins



Reducing the Number  
of Junction Proteins

Gao, Sun, Moore, White, Brink, Mathias  
*J Gen Physiol*, 2011  
137: 507-20.

**“A comprehensive model of lens transport,  
including osmotic  
and hydrostatic pressure gradients in both  
the intracellular  
and extracellular compartments of the lens,  
is beyond the scope of this paper.”**



# Our Contributions

are to Derive Consistent Field Equations  
and Solve Them

**Multiple Fields are Hard to Deal with  
Consistently without Variational Methods**  
*(in engineering models or computations)*

*A great deal of confusion can result  
from nonmathematical reasoning  
Including paradoxes*

**Osmosis through a Semi-permeable Membrane**  
**a Consistent Approach to Interactions**

arXiv:1806.00646



**Shixin Xu**



**Zilong Song**



**Huaxiong Huang**

**A Bidomain Model for Lens Microcirculation**

Biophysical Journal (2019) 116: p. 1171-1184



**Yi Zhu**



**Shixin Xu**

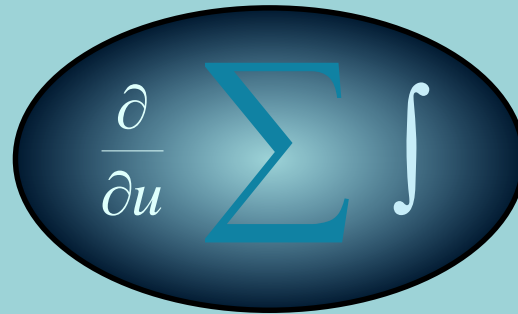


**Huaxiong Huang**

# Inconsistent Models

produce confusion  
even when correct

## Mathematics



replaces  
Inconsistent Models Models  
with **Consistent**  
Partial Differential Equations  
and boundary conditions

# **Scientists and Poets can Reach but Engineers must Grasp**

**and not just reach  
if  
Devices are to Work**  
*Uncalibrated Devices do not Work!*

## **Poets**

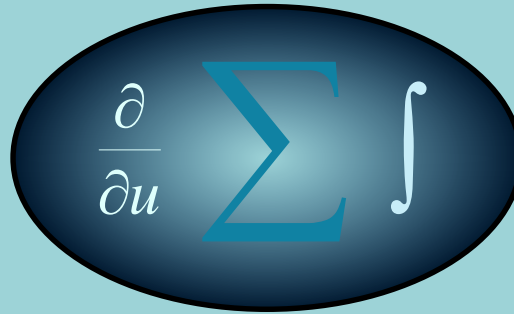
hope we will never learn the difference between dreams and realities

**“Ah, ... a man's reach should exceed his grasp,**

**Or what's a heaven for?”**

**Robert Browning**

*"Andrea del Sarto", line 98*



Mathematics Creates our Standard of Living\*

**Partial Differential Equations**  
**are needed to describe**  
**Flow**  
**driven by multiple fields**

*\*e.g., Electricity, Computers, Fluid Dynamics, Optics, Structural Mechanics, .....*

**We begin at the beginning:**

# **Ionic Solutions are Complex Fluids**

in which

**‘everything interacts with everything else’**

and

**Flows are driven by convection, and diffusion.**

**Shixin Xu, Eisenberg, Song, and Huang**

(2018)

*arXiv:1806.00646, 35 pages.*



# **Bulk Solutions and Membranes are described by Consistent Analysis of Coupled Water and Ionic Diffusion and Flow**

The **Energy Variational Principle and Sharp Boundary Methods** have been applied to a variety of membrane models, allowing **density of solutions to be a function of concentrations**, as is seen every day in chemistry laboratories **apparently for the first time**

Unfortunately, so far, analysis has only been performed for **ideal solutions**.

## **Challenge**

No one seems to know how to formulate coupled water and ion **flow** and diffusion, in the realistic nonideal case found in biology and electrochemistry **NOT equilibrium, not ideal, must deal with saturation and finite size ions**

Shixin Xu, Eisenberg, Song, and Huang (2018)  
*arXiv:1806.00646, 35 pages.*

## **Challenge**

No one seems to know  
how to formulate coupled water and ion **flow** and  
diffusion,  
in the realistic nonideal case found in biology and  
electrochemistry

**NOT equilibrium, not ideal,  
must deal with saturation and finite size ions.**

## **Suggestion:**

**Combine EnVarA with Poisson Fermi approach  
of Jinn Liang Liu,  
*more than anyone else***

Starling's Law has been derived: water flow is proportional to hydrostatic and osmotic difference from the Energy Variational Principle and a Sharp Boundary Methods

**Total Energy Functional.**

$$\begin{aligned}
 E^{\text{tot}} &= E_{kin} + E_{int} + E_{\Gamma} \\
 &= E_{kin} + E_{es} + E_{ion} + E_{\Gamma} \\
 &= \sum_{\pm} \int_{\Omega^{\pm}} (e_{kin}^{\pm} + e_{es}^{\pm} + e_{ion}^{\pm}) dx + \int_{\Gamma} (e_{\Gamma} + \gamma_0) dS \\
 &= \sum_{\pm} \int_{\Omega^{\pm}} \left\{ \frac{1}{2} \rho^{\pm} |\mathbf{u}^{\pm}|^2 + \frac{1}{2} \mathbf{E}^{\pm} \cdot \mathbf{D}^{\pm} + k_B T \sum_i c_i^{\pm} \ln \left( \frac{c_i^{\pm}}{c_0} \right) \right\} dx \\
 &\quad + \int_{\Gamma} \left( \frac{C_m}{2} [\phi]^2 + \gamma_0 \right) dS
 \end{aligned} \tag{3}$$

where  $c_0$  is a characteristic ion density,  $C_m$  is membrane capacitance,  $\gamma_0$  is the membrane surface tension. In the following, square brackets always denote the jumps across the interface.

$\sum_{\pm} \int_{\Omega^{\pm}} f^{\pm} dx = \int_{\Omega^+} f^+ dx + \int_{\Omega^-} f^- dx$  is the sum of integration in left and right compartments weighted by the corresponding value of  $f$ .

The **dissipation functional** is defined as

$$\begin{aligned}
 \Delta &= \sum_{\pm} \int_{\Omega^{\pm}} 2\eta^{\pm} |\mathbf{D}_{\dot{\eta}}^{\pm}|^2 dx + \sum_{\pm} \int_{\Omega^{\pm}} \lambda^{\pm} |\nabla \cdot \mathbf{u}^{\pm}|^2 dx + \frac{1}{k_B T} \sum_{\pm} \int_{\Omega^{\pm}} \sum_i D_i^{\pm} c_i^{\pm} |\nabla \tilde{\mu}_i^{\pm}|^2 dx \\
 &\quad + \int_{\Gamma} G_1([\tilde{\mu}_i]) dS + \int_{\Gamma} G_2(Q_{\rho}) dS
 \end{aligned} \tag{4}$$

Where  $\mathbf{D}_{\dot{\eta}} = (\nabla \mathbf{u} + (\nabla \mathbf{u})^T)/2$  is rate of strain,  $\eta^{\pm}$  and  $\lambda^{\pm}$  are the two Lamé constants<sup>109</sup>,  $\tilde{\mu}_i^{\pm} = \tilde{\mu}_i^{\pm}(c_i^{\pm}, \phi^{\pm}, \rho^{\pm})$  and  $D_i^{\pm}$  are the chemical potential and diffusion coefficient of  $i_{th}$  ion. The first three terms in dissipation functional are the dissipation induced by fluid friction, volume change and ion diffusion in the bulk region. The last two terms are the dissipation induced by irreversible osmosis on the membrane. The forms of  $G_1(x) \geq 0$  and  $G_2(x) \geq 0$ , for any  $x$ , will be discussed later.

**NOT unique**  
**This is a PHYSICAL Model**  
 e.g. it does not include  
**Saturation produced by**  
**Finite size of ions**

**Dissipation IS NOT unique**  
**This is a PHYSICAL Model**  
 e.g. it must it time and  
 concentration dependence and  
 much more  
**BUT**  
**Mathematics must be consistent**  
 with the  
**TOTAL ENERGY FUNCTIONAL**

## Intracellular and Extracellular Velocities (1)

The intracellular velocity depends on the gradients of hydrostatic pressure and osmotic pressure (41,51,62), and the extracellular velocity is determined by the gradients of hydrostatic pressure and electric potential (41,67),

$$u_{ex} = -\frac{\kappa_{ex}}{\mu} \tau_c \frac{d}{dr} P_{ex} - k_e \tau_c \frac{d}{dr} \phi_{ex} \quad (2a)$$

and

$$u_{in} = -\frac{\kappa_{in}}{\mu} \left( \frac{d}{dr} P_{in} - \gamma_m k_B T \frac{d}{dr} O_{in} \right), \quad (2b)$$

where  $\phi_l$  is the electric potential in the  $l$  space,  $\tau_c$  is the tortuosity of extracellular region,  $\mu$  is the viscosity of water,  $k_e$  is introduced to describe the effect of electro-osmotic flow, and  $\kappa_l$  is the permeability of the intracellular region ( $l = in$ ) and the extracellular region ( $l = ex$ ), respectively.

## Flows and Coupling

Then, we obtain the following system for intra- and extracellular velocities in domain  $\Omega = [0, R]$ :

Coupling: 
$$\begin{aligned} & \frac{1}{r^2} \frac{d}{dr} (r^2 \mathcal{M}_{ex} u_{ex}) \\ &= -\mathcal{M}_v L_m (P_{ex} - P_{in} + \gamma_m k_B T (O_{in} - O_{ex})) \end{aligned} \quad (1a)$$

and

Conservation: 
$$\frac{1}{r^2} \frac{d}{dr} (r^2 (\mathcal{M}_{ex} u_{ex} + \mathcal{M}_{in} u_{in})) = 0, \quad (1b)$$

where  $u_l$  and  $P_l$  are the velocity and pressure in the intracellular and extracellular space, respectively, and  $O_l$  is the osmotic pressure with definition

Osmolarities: 
$$O_{ex} = \sum_i C_{ex}^i, \quad O_{in} = \sum_i C_{in}^i + \frac{A_{in}}{V_{in}},$$

where  $C_l^i$  is the concentration of  $i$ th species ion in  $l$  space and  $A_{in}/V_{in}$  is the density of the permanent negatively charged protein. In this work, we assume the permanent negative charged protein is uniformly distributed within intracellular space with a valence of  $\bar{z}$ . Here,  $\mathcal{M}_l$  is the ratio of intracellular area ( $l = in$ ) and extracellular area ( $l = ex$ ),  $\mathcal{M}_v$  is the membrane area per volume unit,  $\gamma_m$  is the intracellular membrane reflectance,  $L_m$  is intracellular membrane hydraulic permeability,  $k_B$  is the Boltzmann constant, and  $T$  is temperature.

## Ion Circulation

With similar assumptions, the conservation of ion concentration yields the following ion flux system:

$$\frac{1}{r^2} \frac{d}{dr} (r^2 \mathcal{M}_{ex} J_{ex}^i) = \mathcal{M}_v j_m^i \quad (4a)$$

and

$$\frac{1}{r^2} \frac{d}{dr} (r^2 (\mathcal{M}_{ex} J_{ex}^i + \mathcal{M}_{in} J_{in}^i)) = 0. \quad (4b)$$

The ion flux in the intracellular region  $J_{in}^i$  and ion flux in the extracellular region  $J_{ex}^i$  are defined as

$$J_{ex}^i = C_{ex}^i u_{ex} - D_{ex}^i \tau_c \frac{d}{dr} C_{ex}^i - D_{ex}^i \tau_c \frac{z^i e}{k_B T} C_{ex}^i \frac{d}{dr} \phi_{ex}, \quad (5a)$$

## Membrane Flux of Ions

$$j_m^i = \frac{g^i}{e z^i} (\phi_{in} - \phi_{ex} - E^i), \quad (6a)$$

$$J_s^i = \frac{G^i}{e z^i} (\phi_{in} - \phi_{ex} - E^i), \quad (6b)$$

where  $E^i = k_B T / e z^i \log(C_{ex}^i / C_{in}^i)$  is the Nernst potential (an expression of the difference of chemical potential) of the  $i$ th species ion.

# **Electroneutral Expansion**

from PNP = Poisson Nernst Planck,  
named by Eisenberg & Chen, 1993, Biophysical Journal **64**: A22.

## NOT electroneutral

Because of the capacitance of the cell membrane, assumptions of exact charge neutrality can easily lead to paradoxes because they oversimplify Maxwell's equations by leaving out altogether the essential role of charge. We use the analysis of (70) and thus introduce a linear correction term replacing the charge neutrality condition (41,51) without introducing significant error (see also (71)),

$$(1 - \eta) \left( \sum_i e z^i C_{ex}^i \right) = -\mathcal{M}_v C_m (\phi_{in} - \phi_{ex}) \quad (10a)$$

and

$$\eta \left( \sum_i e z^i C_{in}^i + \bar{z} e \frac{A_{in}}{V_{in}} \right) = \mathcal{M}_v C_m (\phi_{in} - \phi_{ex}), \quad (10b)$$

where  $\eta$  is the porosity of intracellular region and  $C_m$  is capacitance per unit area.

# Simplified Model

by well-defined  
asymptotic expansion

With ordering

$$\{\delta_1, \delta_8\} \subset O(\varepsilon), \quad \{\delta_0, \delta_3\} \subset O(\varepsilon^2), \\ \{\delta_2, \delta_4, \delta_5, \delta_6, \delta_7\} \subset o(\varepsilon^2). \quad (16)$$

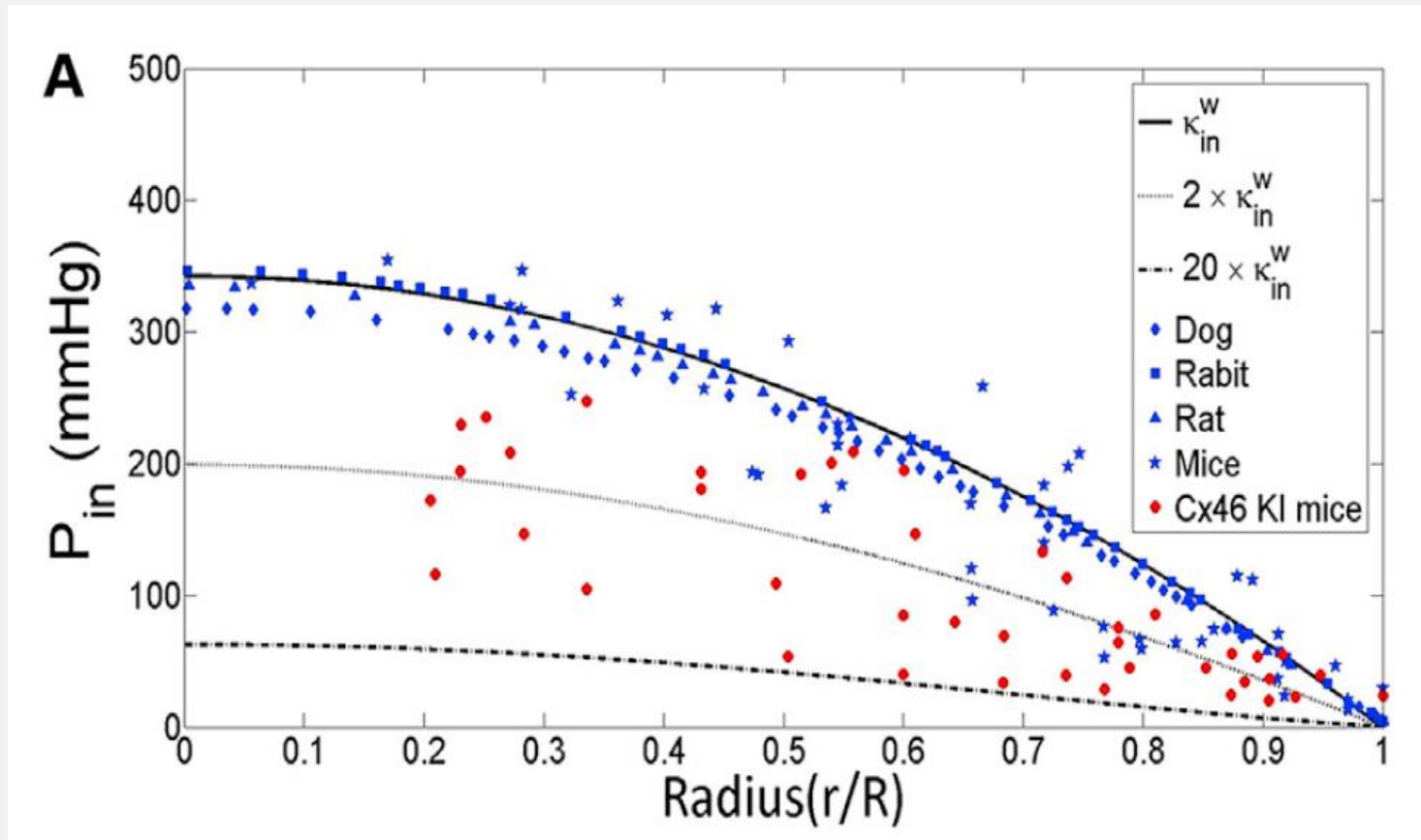
$$\delta_9 = D_l^{Cl} - D_l^K \text{ and } \delta_{10} = D_l^{Cl} - D_l^{Na}, l = in, ex, \text{ it yields} \\ \delta_9 = O(\varepsilon^2), \quad \delta_{10} = O(\varepsilon). \quad (17)$$

$$\delta_0 = \frac{\mathcal{M}_{ex}}{\mathcal{M}_{in}}, \\ \delta_1 = \frac{k_e \tau_c k_B T}{e R u_{ex}^*}, \\ \delta_2 = \frac{\mu R u_{in}^*}{\kappa_{in} \gamma_m k_B T O^*}, \\ \delta_3 = \frac{P^*}{\gamma_m k_B T O^*}, \\ \delta_4 = \frac{\mathcal{M}_{in} u_{in}^*}{R \mathcal{M}_v L_m \gamma_m k_B T O^*}, \\ \delta_5 = \frac{u_{in}^*}{L_s \gamma_s k_B T O^*}, \\ \delta_6 = \frac{\mathcal{M}_v C_m k_B T}{e^2 C^* \eta},$$

$$\delta_7 = \frac{\mathcal{M}_v C_m k_B T}{e^2 C^* (1 - \eta)}, \\ \delta_8 = \frac{\mathcal{M}_{ex} D_{ex}^*}{\mathcal{M}_{in} D_{in}^*}, \\ \delta_9 = \frac{D_l^{Cl} - D_l^K}{D_l^*}, l \in \{in, ex\} \\ \delta_{10} = \frac{D_l^{Cl} - D_l^{Na}}{D_l^*}, l \in \{in, ex\} \\ \delta_{11} = \frac{C_{in}^{Cl,0}}{C_{ex}^{Cl,0}},$$



# Comparison with Experimental Results



# Dependence on 'Connexin Permeability' $\kappa_{in}$

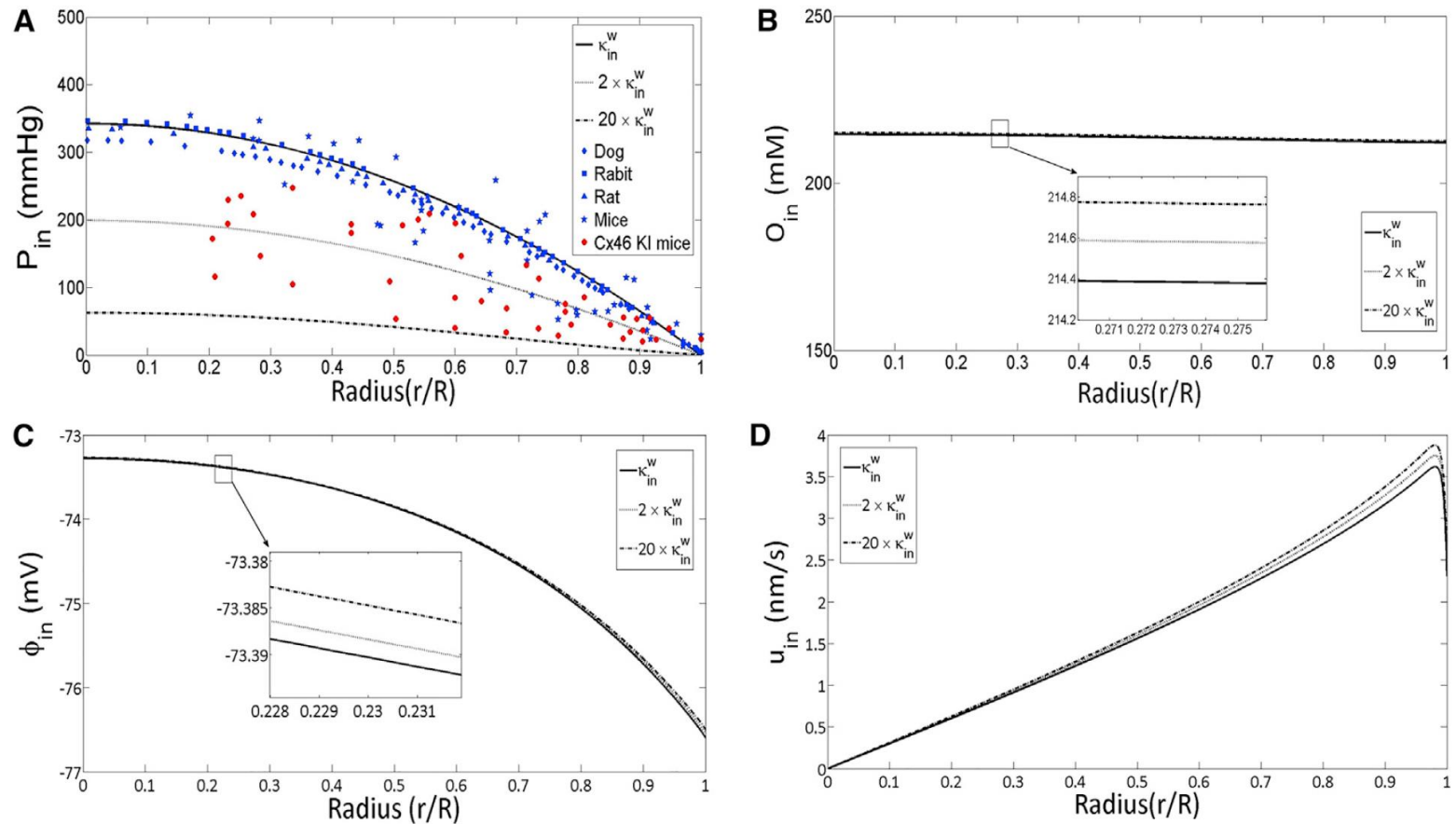
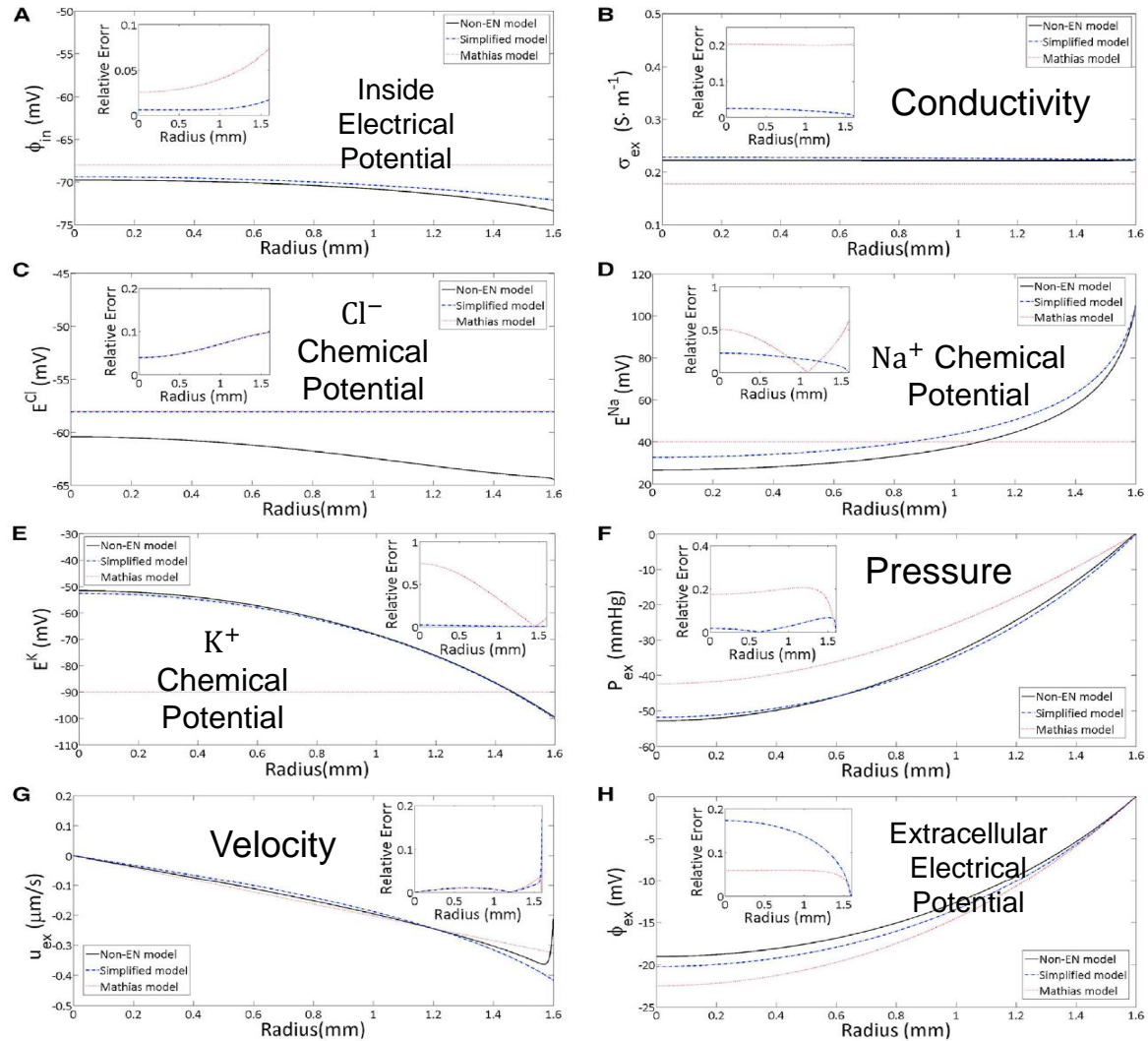


FIGURE 3 Comparison between different  $\kappa_{in}$ . (A) Comparison between simulation results and experimental results. The experimental data of dog, rabbit, and rat come from (47). Mice and Cx46 KI mice come from (28). According to (28), the Cx46 KI mice lens has twice the number density of lens gap-junction channels compared to mice. The parameter  $\kappa_{in}^w = 4.6830 \times 10^{-20}/m^2$ , and radius is written in dimensionless units for different species. (B) Space distribution of intracellular osmotic pressure ( $O_{in}$ ). (C) Space distribution of intracellular electric potential ( $\phi_{in}$ ). (D) Space distribution of intracellular water velocity ( $u_{in}$ ). To see this figure in color, go online.

# Non-Electroneutral Model vs. Simplified, and Mathias Models



# **Resolution of Troubling Paradoxes in our Models**

Perturbation Expansion shows that  
**Many intracellular and extracellular quantities  
are INSENSITIVE  
to increases in permeability  
EVEN BY A FACTOR of 20×**

Because many quantities  
**depend only on the extracellular potential,**  
and the  
Extracellular Potential is Insensitive  
to  
Permeability

## Effect of Permeability $\kappa_{in}$

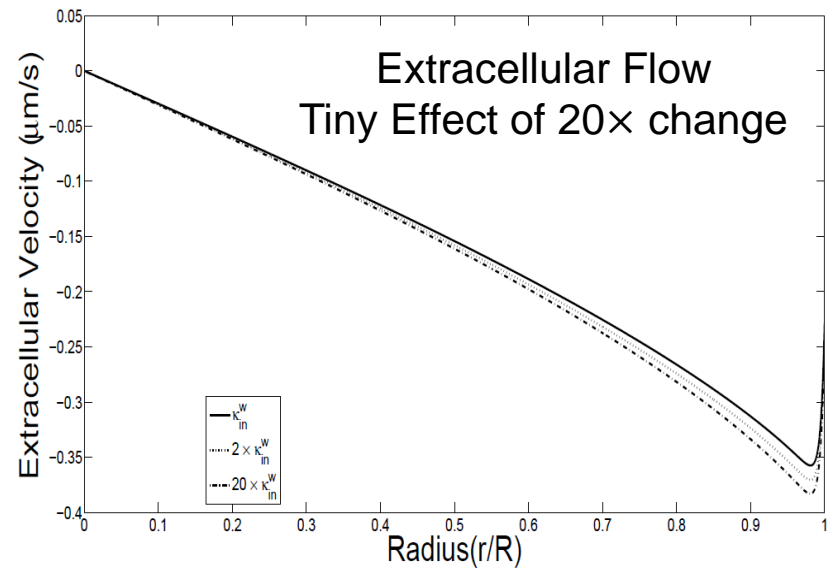
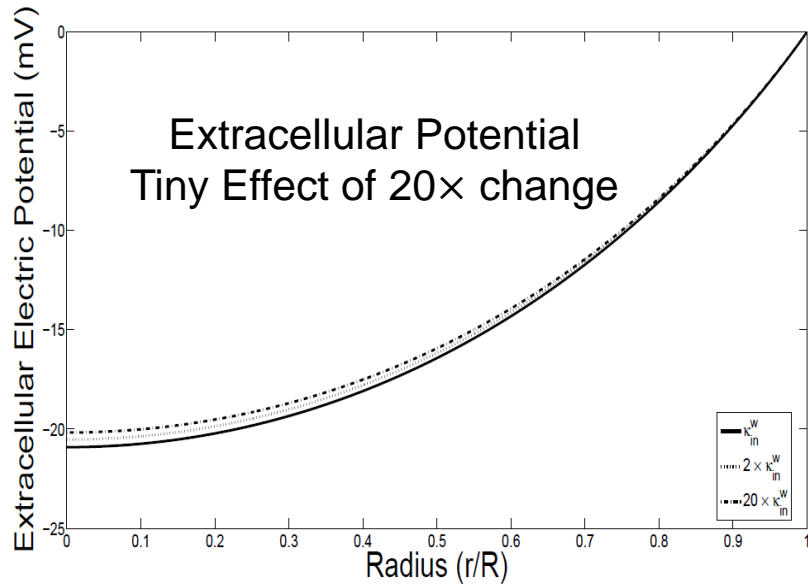
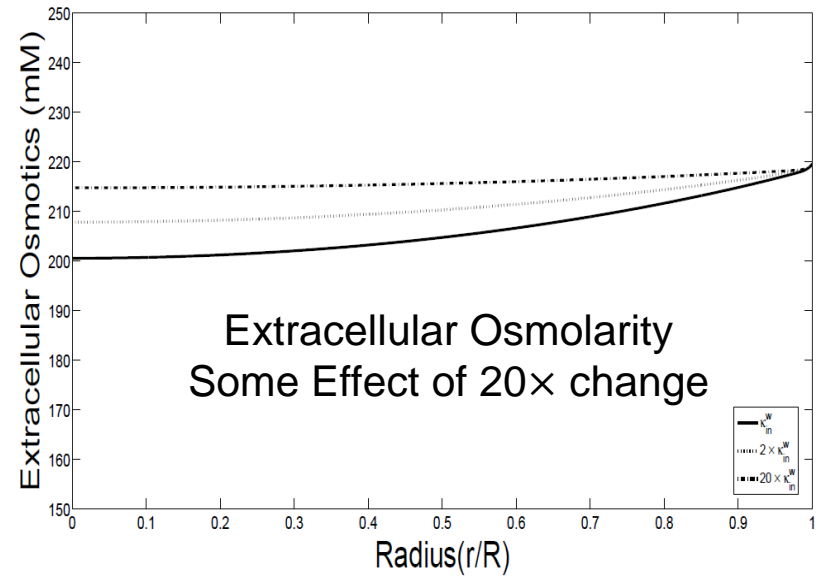
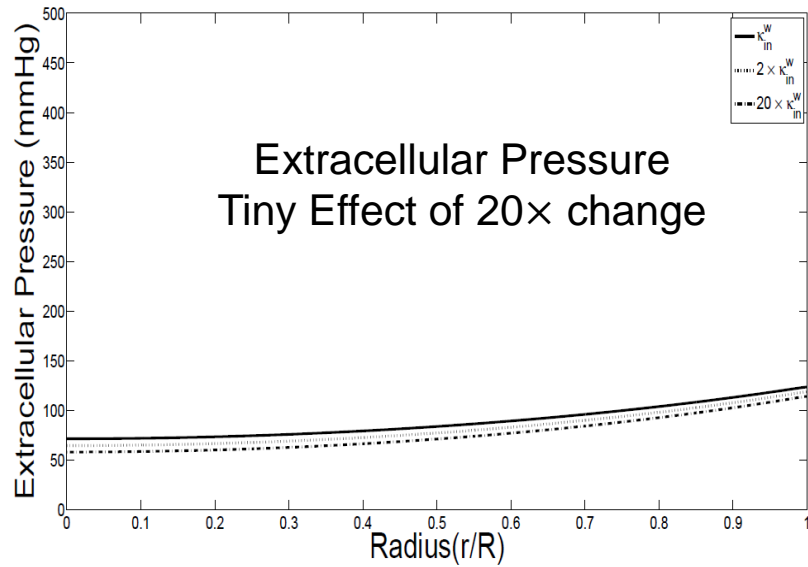
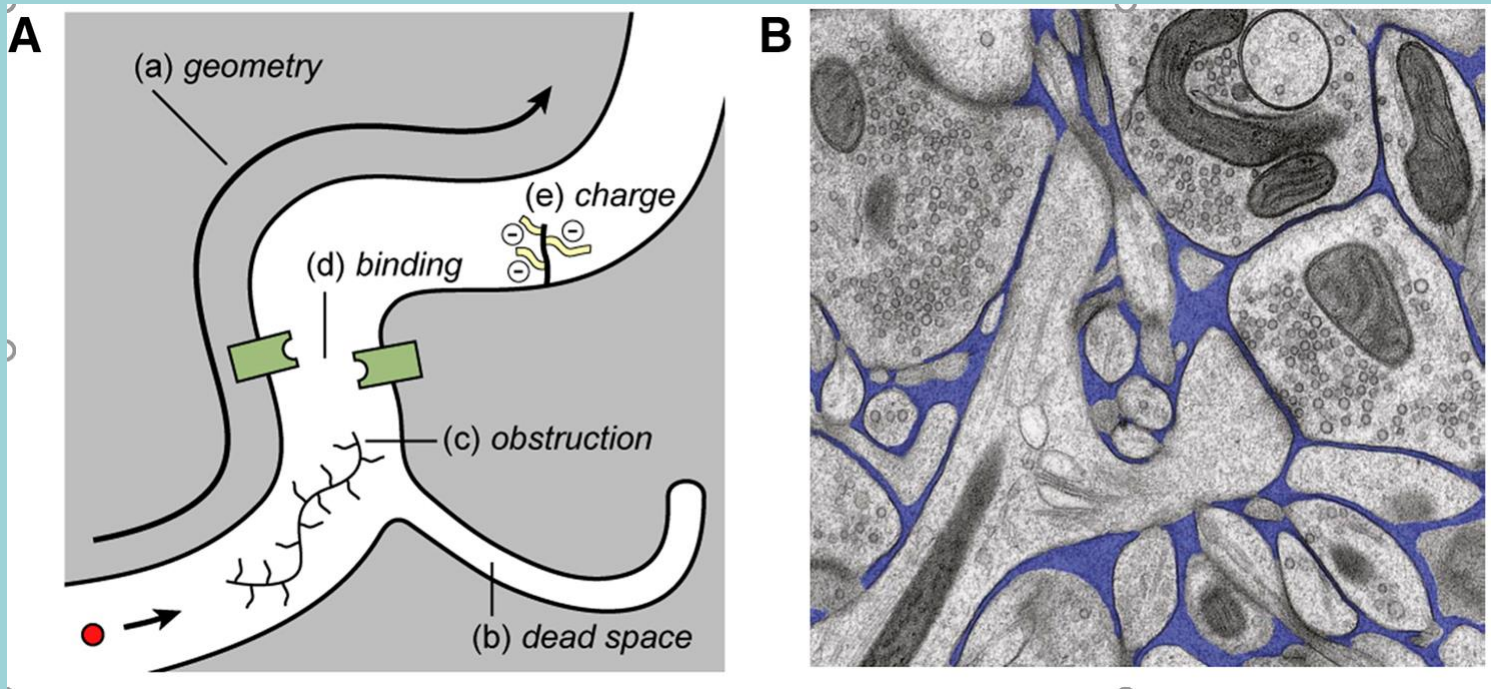


Figure D.5: Comparison between different  $\kappa_{in}$ .

What next?

## “Brain Extracellular Space: The Final Frontier”

Nicholson and Hrabětová,  
*Biophysical Journal*, 2017. 113: p. 2133-2142.



**Stirred by an Osmotic Pump  
like the lens,  
at work and in sleep ??**

*Any Questions ?*