

Figure Legends

1. The geometry of the model RyR pore. In the experiments and calculations, the lumen of the sarcoplasmic reticulum (SR) is electrically grounded. The circle around each labeled amino acid is meant to illustrate the range of the motion of the terminal carboxyl group. Aspartates (solid circles) are given a radius of 5 Å and glutamates (dashed circles) 7 Å. Only the amino acids of one of the four identical RyR subunits is shown. Asp-4945, Asp-4938, Asp-4899, Glu-4900, and Glu-4902 are the only amino acids explicitly modeled in the theory. The GGGIG sequence (4894-4898 in the numbering) at the cytosolic end of the selectivity filter is only a reference point for readers familiar with the RyR sequence.
2. Experimental verification of the AMFE predicted by the theory. The lines are the theory and the symbols are the experimental data with standard error bars and the number of experiments in parentheses. The solid line and symbols are the addition of Ca^{2+} to Na^+ and the dashed line and open symbols are the addition of Ca^{2+} to Cs^+ . In all cases, the monovalent-chloride concentration was 150 mM in both cytosolic and luminal baths and the indicated amount Ca^{2+} was added to the luminal bath. The current at -20 mV is shown.
3. The partitioning $\ln(\rho_i(x)/\rho_i^{\text{bath}})$ of K^+ (panel A) and Ca^{2+} (panel B) from the bath into the pore. $[\text{K}^+] = 150$ mM and the indicated $[\text{Ca}^{2+}]$ is in both baths.
4. The electrostatic component of partitioning $z_i e \phi(x)/kT$ of K^+ (panel A) and Ca^{2+} (panel B) in the pore. $[\text{K}^+] = 150$ mM and the indicated $[\text{Ca}^{2+}]$ is in both baths.
5. The screening component of partitioning $\Delta\mu_i^{\text{SC}}(x)/kT$ of K^+ (solid lines) and Ca^{2+} (dashed lines) in the pore. $[\text{K}^+] = 150$ mM and $[\text{Ca}^{2+}]$ is changed from 1 μM to 50 mM. Because the curves are so close together $[\text{Ca}^{2+}]$ is not indicated.
6. The excluded-volume (hard-sphere) component of partitioning $\Delta\mu_i^{\text{HS}}(x)/kT$ of K^+ (solid lines) and Ca^{2+} (dashed lines) in the pore. $[\text{K}^+] = 150$ mM and $[\text{Ca}^{2+}]$ is changed from 1 μM to 50 mM. Because the curves are so close together $[\text{Ca}^{2+}]$ is not indicated.
7. Components of the binding selectivity from Eq. (6) in the selectivity filter ($x = 20$ Å in Fig. 1 that shows the geometry of the pore). $[\text{K}^+] = 150$ mM and the indicated $[\text{Ca}^{2+}]$ is in both baths. The horizontal-hatched bar is the number advantage, the diagonal-hatched bar is the mean electrostatic advantage, the cross-hatched bar is the screening advantage, and the solid bar is the excluded-volume advantage. The horizontal line is the binding selectivity of Eq. (6) (i.e., the sum of all the terms). A positive term favors the binding of Ca^{2+} while a negative term favors K^+ .
8. Concentration profiles in the pore of the monovalent cation (panel A) and Ca^{2+} (panel B). For each indicated monovalent cation X^+ , $[\text{X}^+] = 150$ mM and $[\text{Ca}^{2+}] = 1$ mM in both baths.
9. Components of the binding selectivity from Eq. (6) in the selectivity filter ($x = 20$ Å in Fig. 1 that shows the geometry of the pore). For each indicated monovalent cation X^+ , $[\text{X}^+] = 150$ mM and $[\text{Ca}^{2+}] = 1$ mM in both baths. Ion diameters: Li^+ 1.33 Å; Na^+ 2.00 Å; K^+ 2.76 Å; Cs^+ 3.42 Å. The horizontal-hatched bar is the number advantage, the diagonal-hatched bar is the mean electrostatic advantage,

- the cross-hatched bar is the screening advantage, and the solid bar is the excluded-volume advantage. The horizontal line is the binding selectivity of Eq. (6) (i.e., the sum of all the terms). A positive term favors the binding of Ca^{2+} while a negative term favors K^+ .
10. Concentration profiles in the pore of K^+ (panel A) and Ca^{2+} (panel B) for the native (WT) channel (solid line) and the mutation D4899N (dashed line). $[\text{K}^+] = 150 \text{ mM}$ and $[\text{Ca}^{2+}] = 1 \text{ mM}$ in both baths. In the model, the mutation is created by “turning off” the charge on the four Asp-4899. The mutation site is the region from which the charge on the four Asp-4899 has been removed.
 11. Concentration profiles in the pore of K^+ (panel A) and Ca^{2+} (panel B) for the native (WT) channel (solid line) and the mutation D4938N (dashed line). $[\text{K}^+] = 150 \text{ mM}$ and $[\text{Ca}^{2+}] = 1 \text{ mM}$ in both baths. In the model, the mutation is created by “turning off” the charge on the four Asp-4938. The mutation site is the region from which the charge on the four Asp-4938 has been removed.
 12. Profiles of the binding selectivity from Eq. (6) (panel A) and its components (excluded volume, panel B; mean electrostatic, panel C; screening, panel D) for the native (WT) channel (solid line) and the mutation D4899N (dashed line). The conditions are those described in Fig. 9.
 13. Profiles of the binding selectivity from Eq. (6) (panel A) and its components (excluded volume, panel B; mean electrostatic, panel C; screening, panel D) for the native (WT) channel (solid line) and the mutation D4938N (dashed line). The conditions are those described in Fig. 10.
 14. Current/voltage curves in KCl (A-H). (I) The conductance at reversal potential with cytosolic $[\text{K}^+]$ is held at 250 mM and luminal $[\text{K}^+]$ is varied. For both experiment and theory the current/voltage was fitted with a line and the slope is plotted. In this and the following figures, concentrations are listed as cytosolic | luminal. The solid lines are the model and symbols are the experimental data.
 15. Current/voltage curves of (A) the D4899N (■) and E4900Q (□) mutants and (B) the D4938N mutant in 250 mM symmetric KCl.
 16. Current/voltage curves in LiCl. The dashed line is the model result for 250 mM cytosolic and 25 mM luminal bath concentrations (Δ).
 17. Current/voltage curves in NaCl.
 18. Current/voltage curves in RbCl.
 19. Current/voltage curves in CsCl.
 20. Mole fraction experiments at 250 mM total cation concentration in symmetric solutions. (A) NaCl and CsCl mixtures. The experimental point at mole fraction 0.6 is statistically significantly different than the point at mole fraction 1 ($p < 0.05$). (B) LiCl and KCl mixtures.
 21. Current/voltage curves in bi-ionic conditions.
 22. Current/voltage curves with divalent and monovalent cations. (A) KCl and CaCl_2 . (B) NaCl and CaCl_2 . (C) CsCl and CaCl_2 . (D) KCl and MgCl_2 . In both baths are 250 mM monovalent-Cl and in the luminal bath is 5 mM (■), 10 mM (□), and 50 mM (▲) divalent- Cl_2 or the baths contain 250 mM cytosolic monovalent-Cl and 25 mM luminal divalent- Cl_2 (Δ). Current/voltage curves of (E) the D4899N (■) and E4900Q (□) mutants and (F) the D4938N mutant in 250 mM symmetric KCl and 10 mM luminal CaCl_2 .

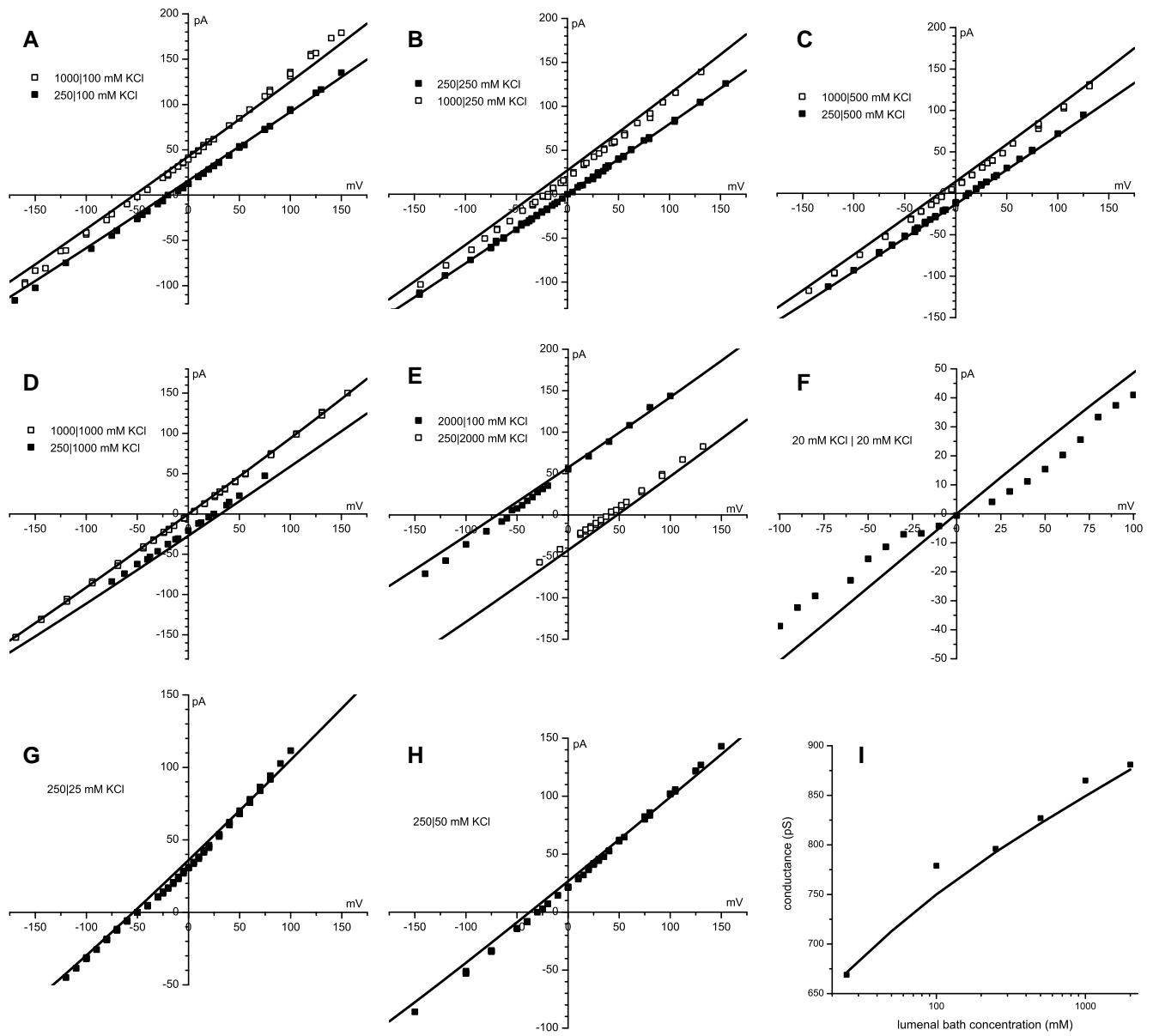


Fig. 14

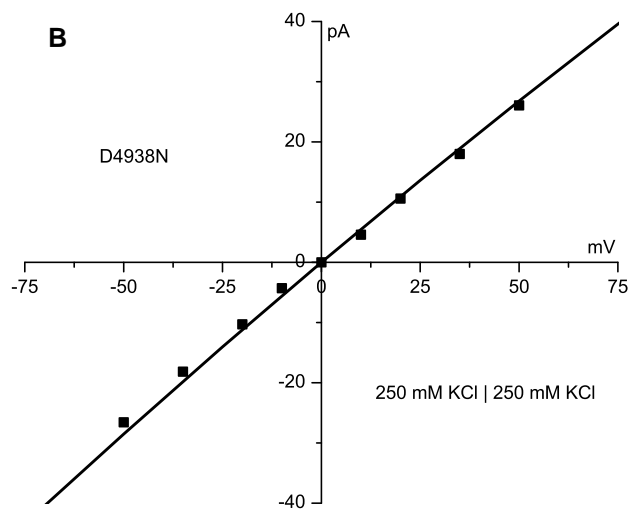
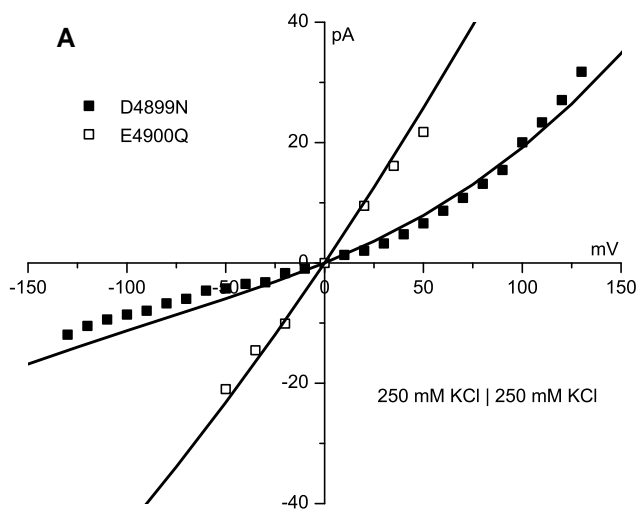


Fig. 15

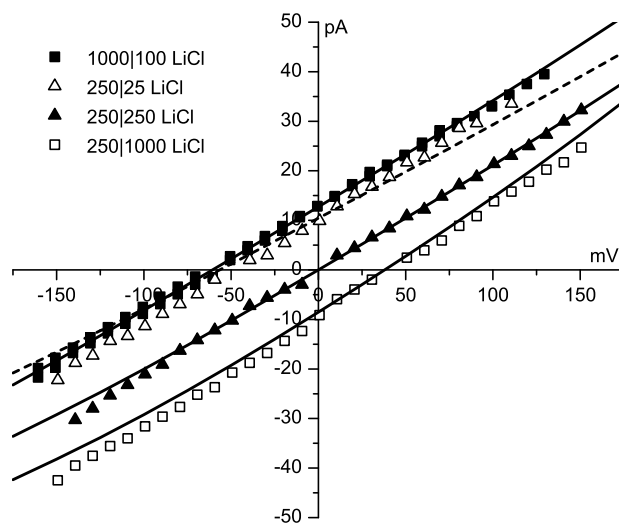


Fig. 16

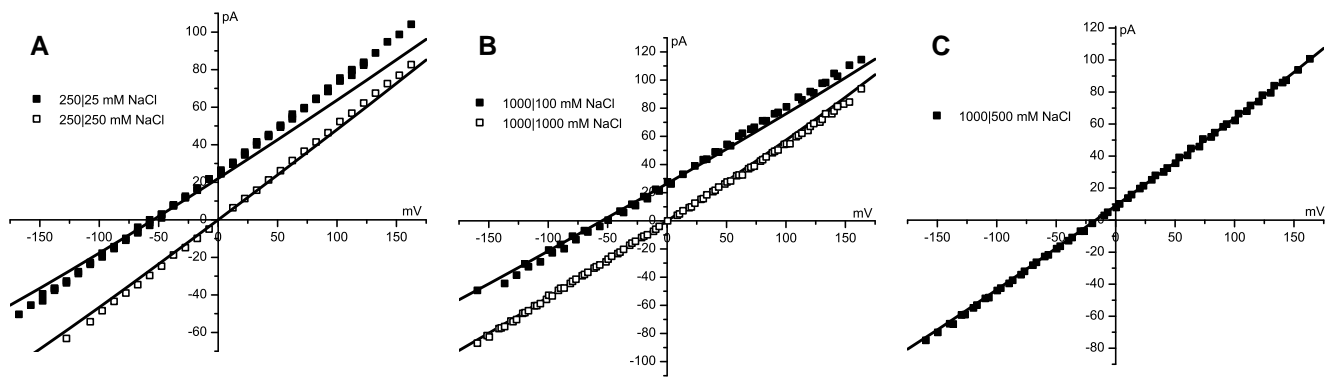


Fig. 17

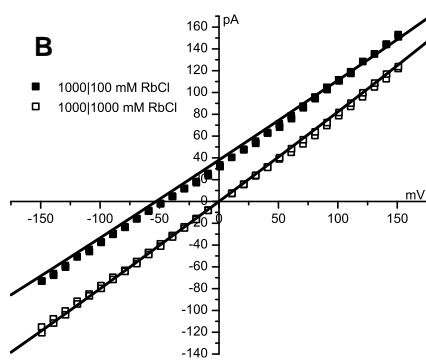
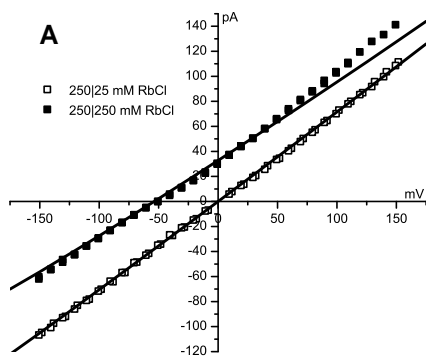


Fig. 18

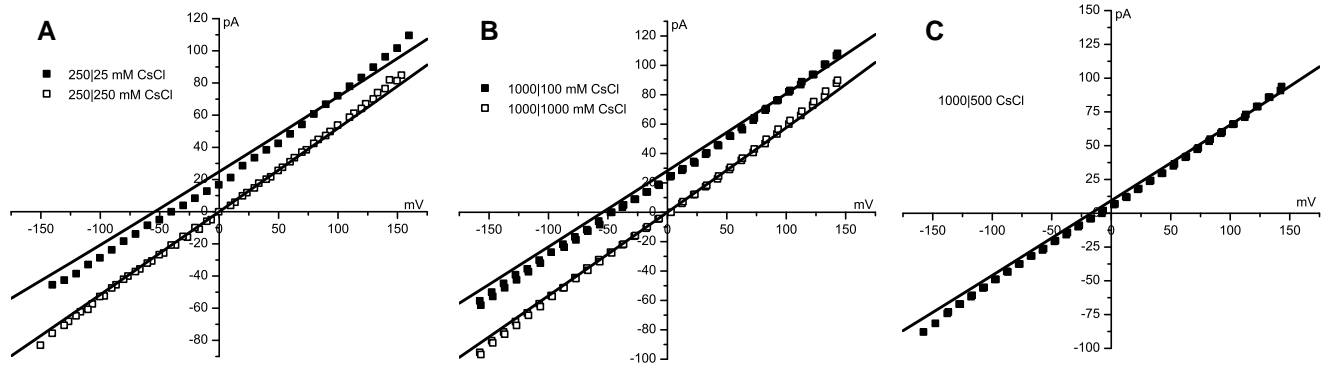


Fig. 19

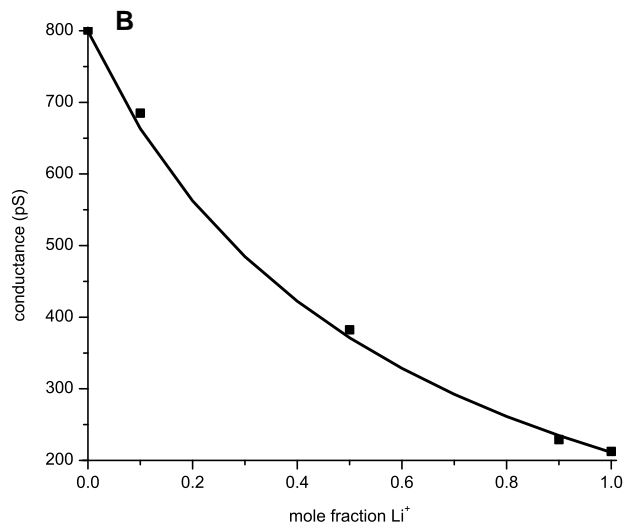
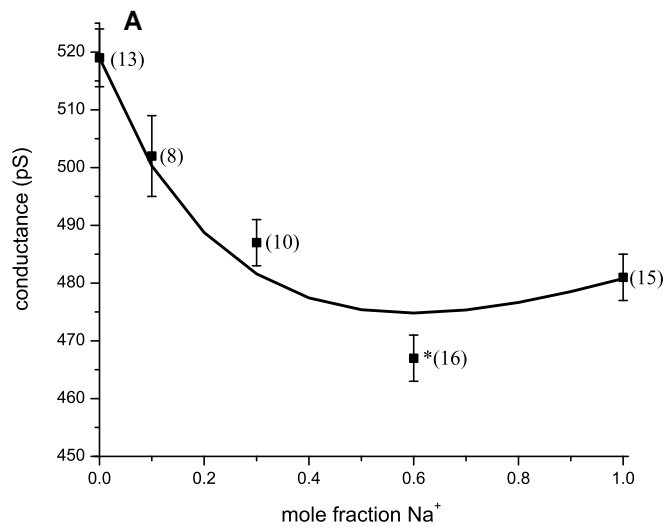


Fig. 20

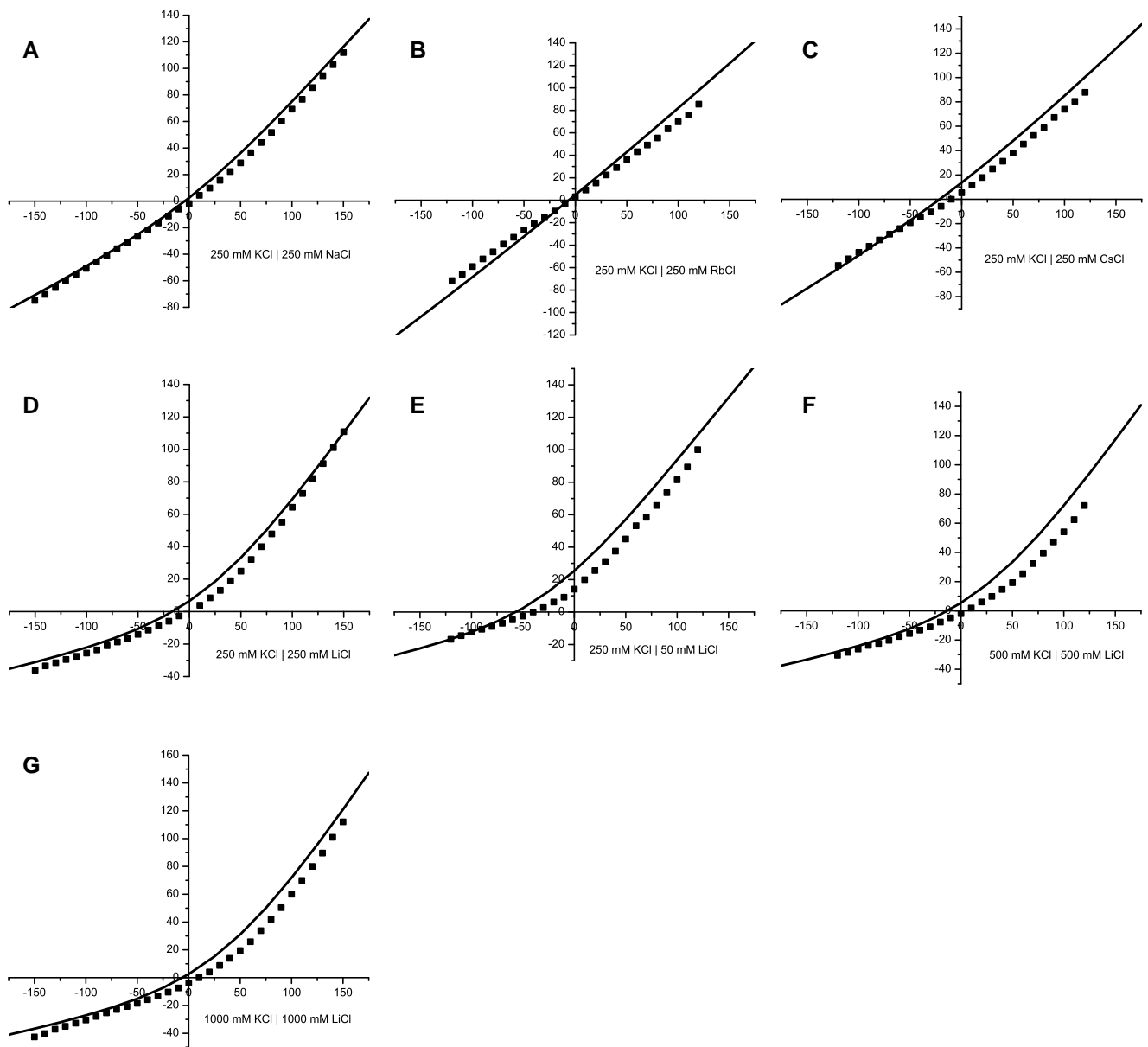


Fig. 21

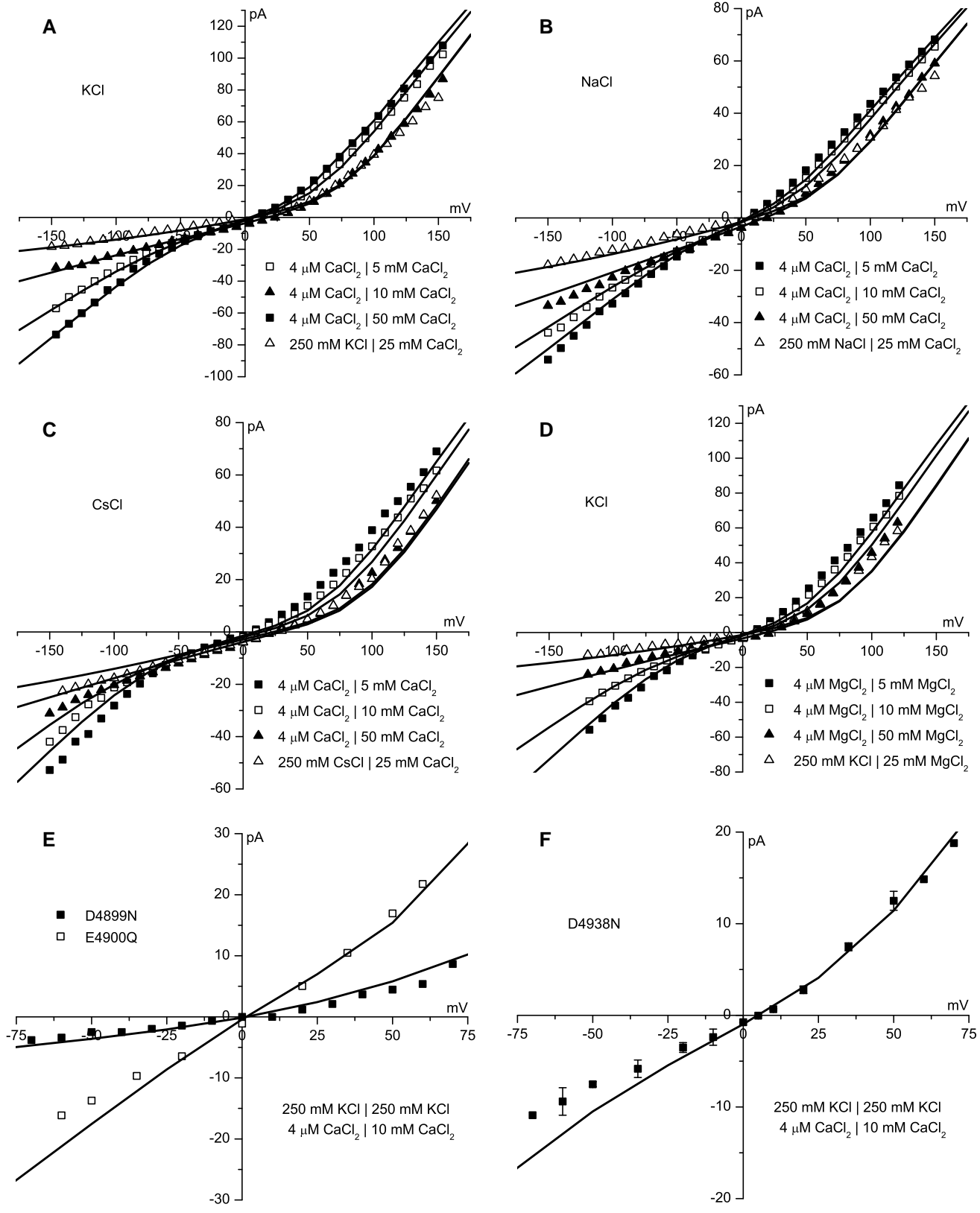


Fig. 22